

The application of ^{19}F NMR spectroscopy in biomolecular studies as a potent technique for structural analysis of the proteins and protein-protein complexes in solution.

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The application of ^{19}F NMR spectroscopy in structural studies of proteins has emerged as a potent tool within the other methods of structural biology. While the classical spectroscopic methods, such as ^1H , ^{13}C , and ^{15}N NMR, routinely used to structural analysis proteins in solution, the utilization of ^{19}F NMR gain couple of new possibilities to contribute in structural studies. Specifically, the 100% naturally abundant of the ^{19}F nucleus, making it an ideal candidate for NMR analysis. With a spin of $1/2$ and a high gyromagnetic ratio, ^{19}F isotope exhibits exceptional sensitivity, approximately 83% that of ^1H . The shielding of the ^{19}F nucleus is predominantly governed by a substantial chemical shift anisotropy (CSA). Consequently, fluorine chemical shifts are exquisitely responsive to variations in the local molecular environment, boasting a chemical shift range nearly 100 times larger than that of ^1H . Another advantage of employing ^{19}F as an NMR probe lies in its virtual absence from the majority of naturally occurring biomolecules. This characteristic allows for the investigation of fluorinated proteins in a wide range of routinely used buffer systems and environments, without interference from background signals. Furthermore, the van der Waals radius of the ^{19}F atom, measuring 1.47 \AA , positions it between the VdW radii of hydrogen (1.2 \AA) and oxygen (1.52 \AA) suggesting that incorporating ^{19}F instead of ^1H nuclei have minimal perturbing effects and often exerts little influence on a protein's biological activity.

In the Laboratory of Biological NMR, we perform the synthesis and purification of the protein, containing fluorinated versions of aromatic residues - tryptophan, phenylalanine, and tyrosine. Based on these achievements, we perform several research projects focused on the structural analysis of the proteins and protein-protein complexes utilizing ^{19}F NMR spectroscopy. Our recent experimental findings affirm that the incorporation of fluorine atoms into aromatic residues makes it possible to obtain valuable experimental data about localization structural modifications, ligand binding, conformational dynamics, and protein interactions at the atomic scale. At the moment, the ^{19}F NMR spectroscopy seems to be a promising and cost-effective technique, which substantially increases our knowledge about complex biomolecular systems.