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Normal tissue injury induced by photon and proton therapies

Mechanisms, Gaps and Opportunities

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Outline

- ✓ **Proton vs. photon therapy data for normal tissue injury (clinical trials)**
- ✓ **Impact of normal tissue response on radiotherapy process optimization – mechanisms**
- ✓ **Normal tissue response – pre-clinical studies**
- ✓ **Gaps and opportunities**
- ✓ **Future directions**



Due to physical properties, protons and heavy ions deposit energy more selectively than X-rays, allowing a higher local control of the tumor.

Thus, the damage induced in normal tissues surrounding the tumor is limited.

The general aim for radiotherapy is to **assist optimisation of cancer cells killing** while **minimising harmful effects to normal tissue of individual patients.**

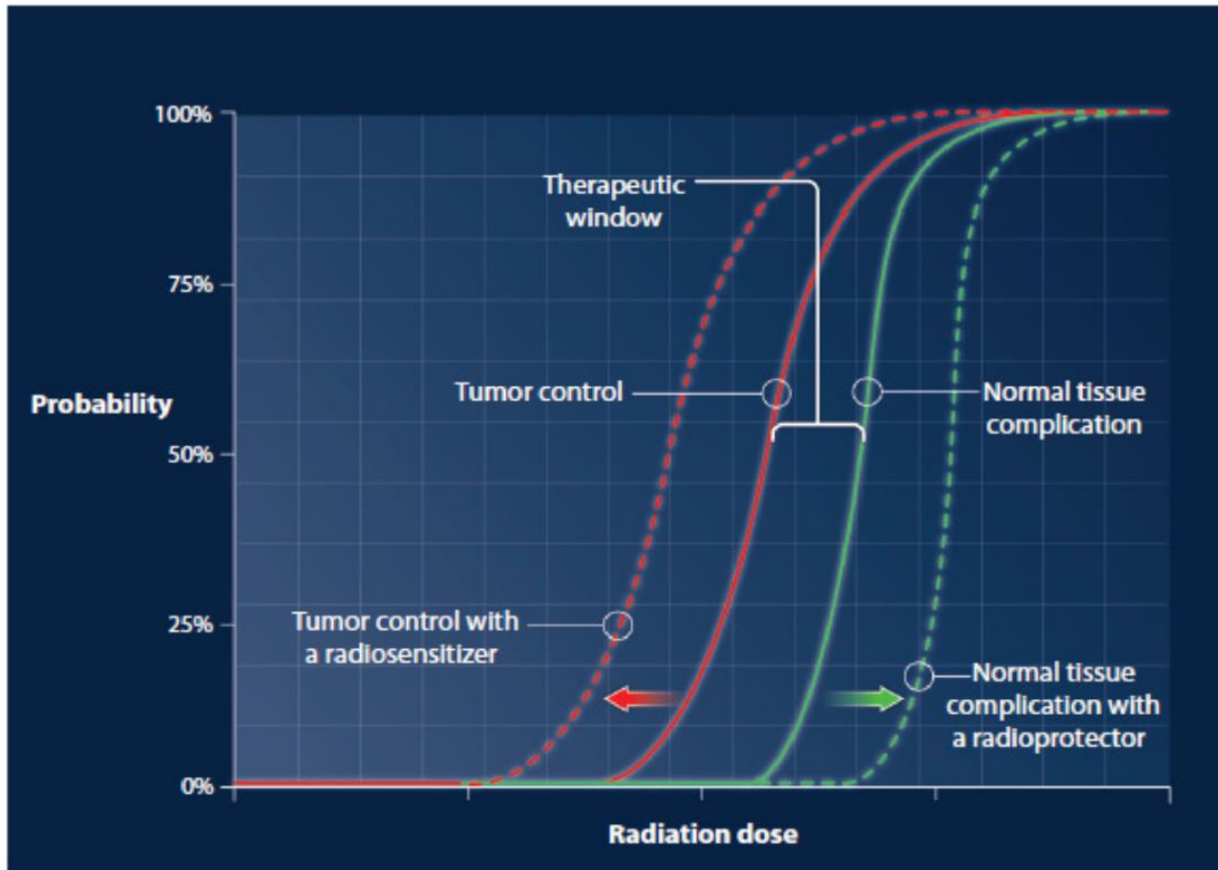


Fig. 1. An idealized graphical representation of tissue effects vs. radiation dose.



The **tumor** and the surrounding **microenvironment** are closely related and interact constantly.

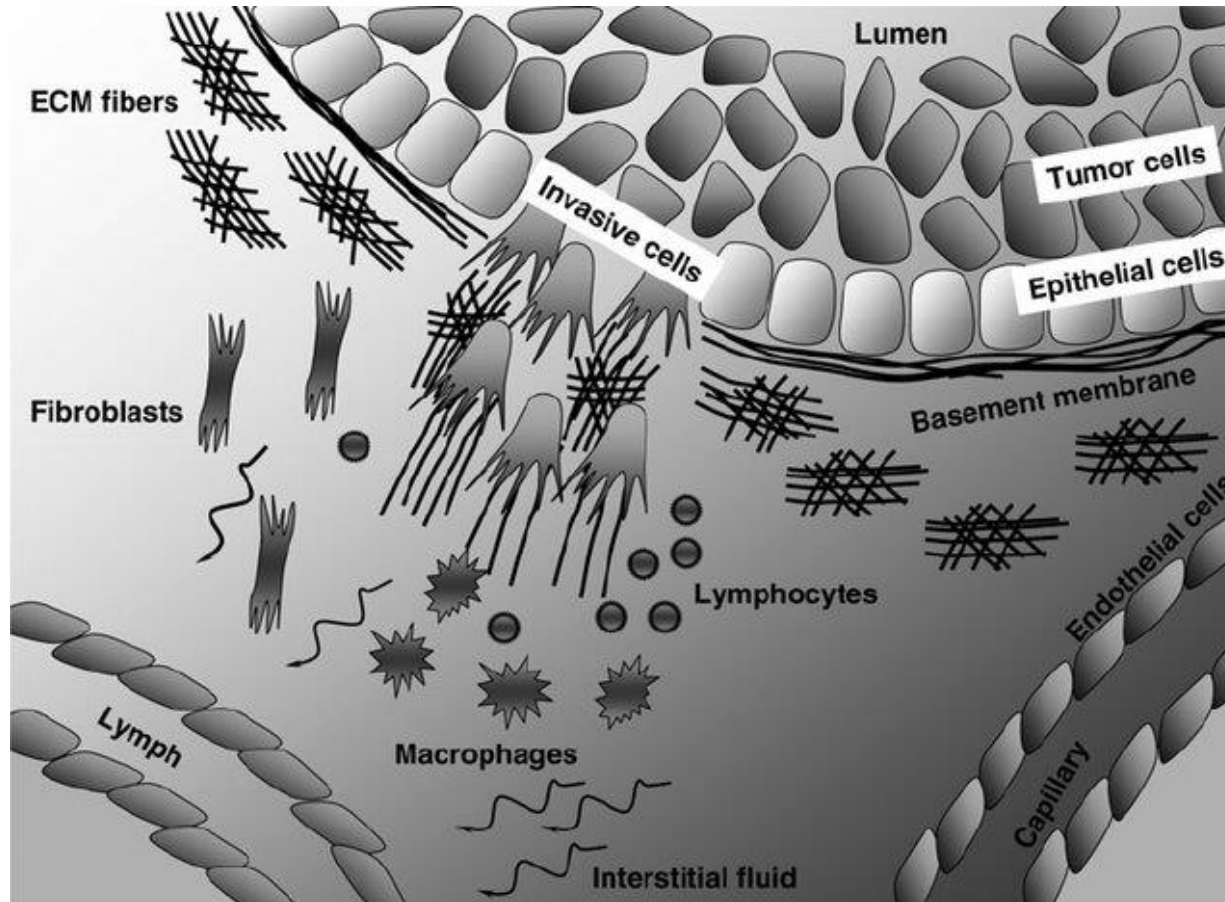


Fig. 2. A scheme of tumor microenvironment components.



CRITICAL REVIEW | ARTICLES IN PRESS

Normal Tissue Injury Induced by Photon and Proton Therapies: Gaps and Opportunities

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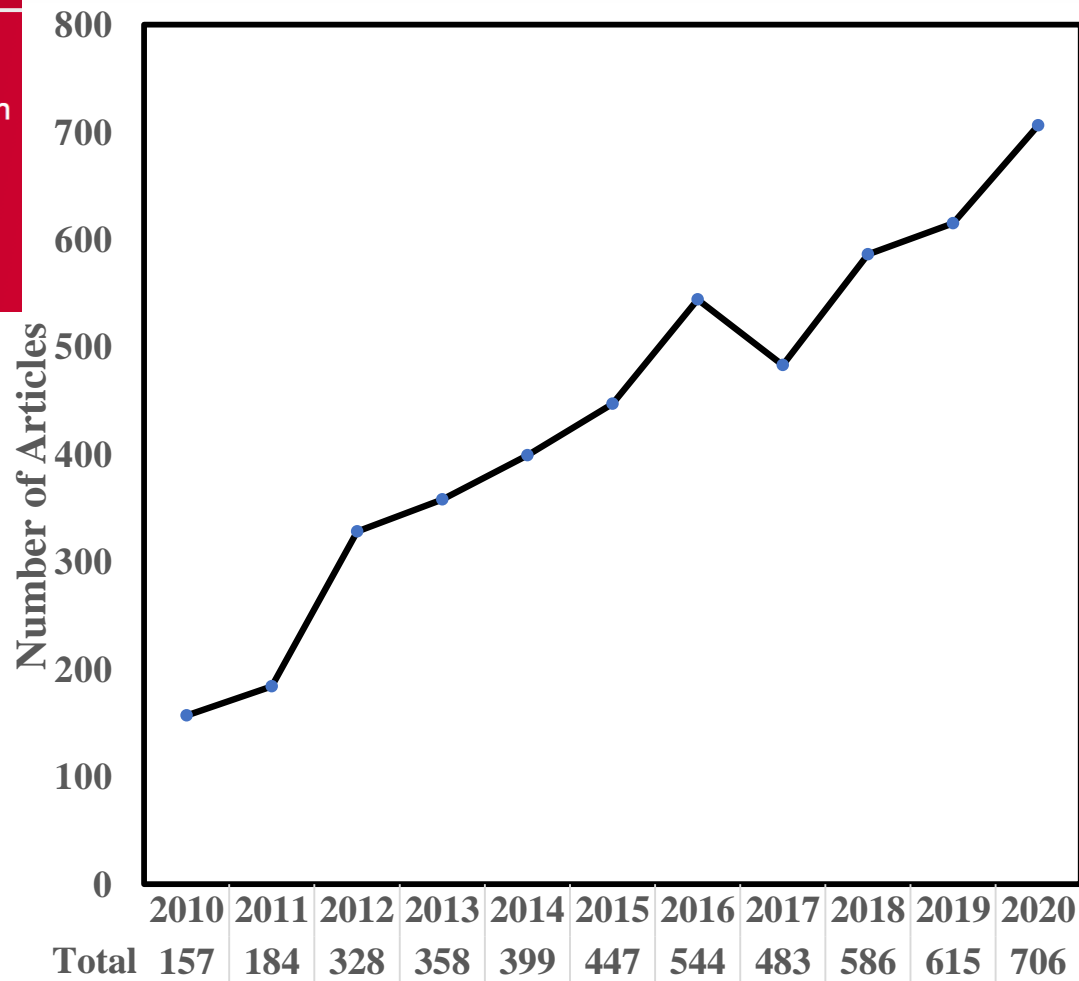


Fig. 3. Number of articles published in PT during the last decade.



Article Type	Total Publications	Publications/Year	Citations/Year		
			Max	Mean	SEM
All Articles	4807	437	51.88	2.32	0.05

According to the latest updates (2020) of the Particle Therapy Co-Operative Group (PTCOG),

- 110 particle therapy facilities** – in operation
- 37 under construction
- 28 in various stages of planning

More than **220,000 patients** have already undergone PT treatments.

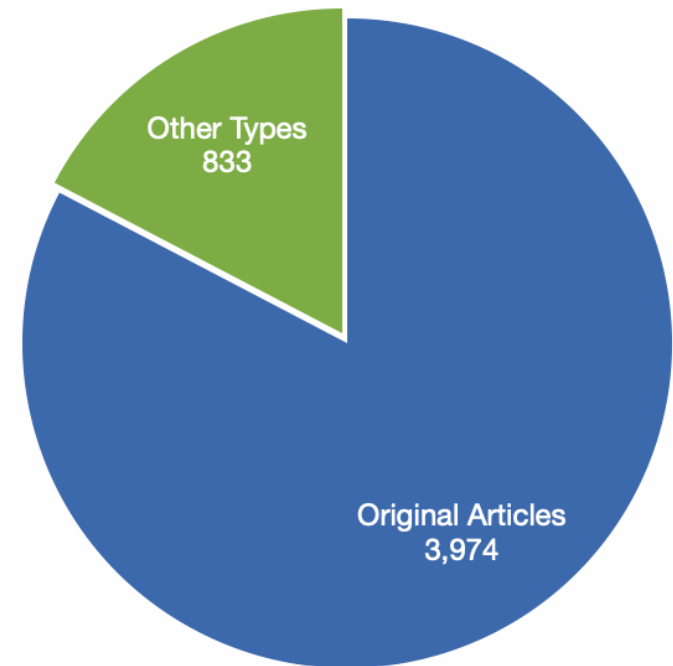


Fig. 4. Summary statistics of PT published articles.

<https://www.ptcog.ch/index.php/other-news>



U.S. National Library of Medicine's
ClinicalTrials.gov
(<https://clinicaltrials.gov>)

Fig. 5. Details of clinical trials conducted with PT.

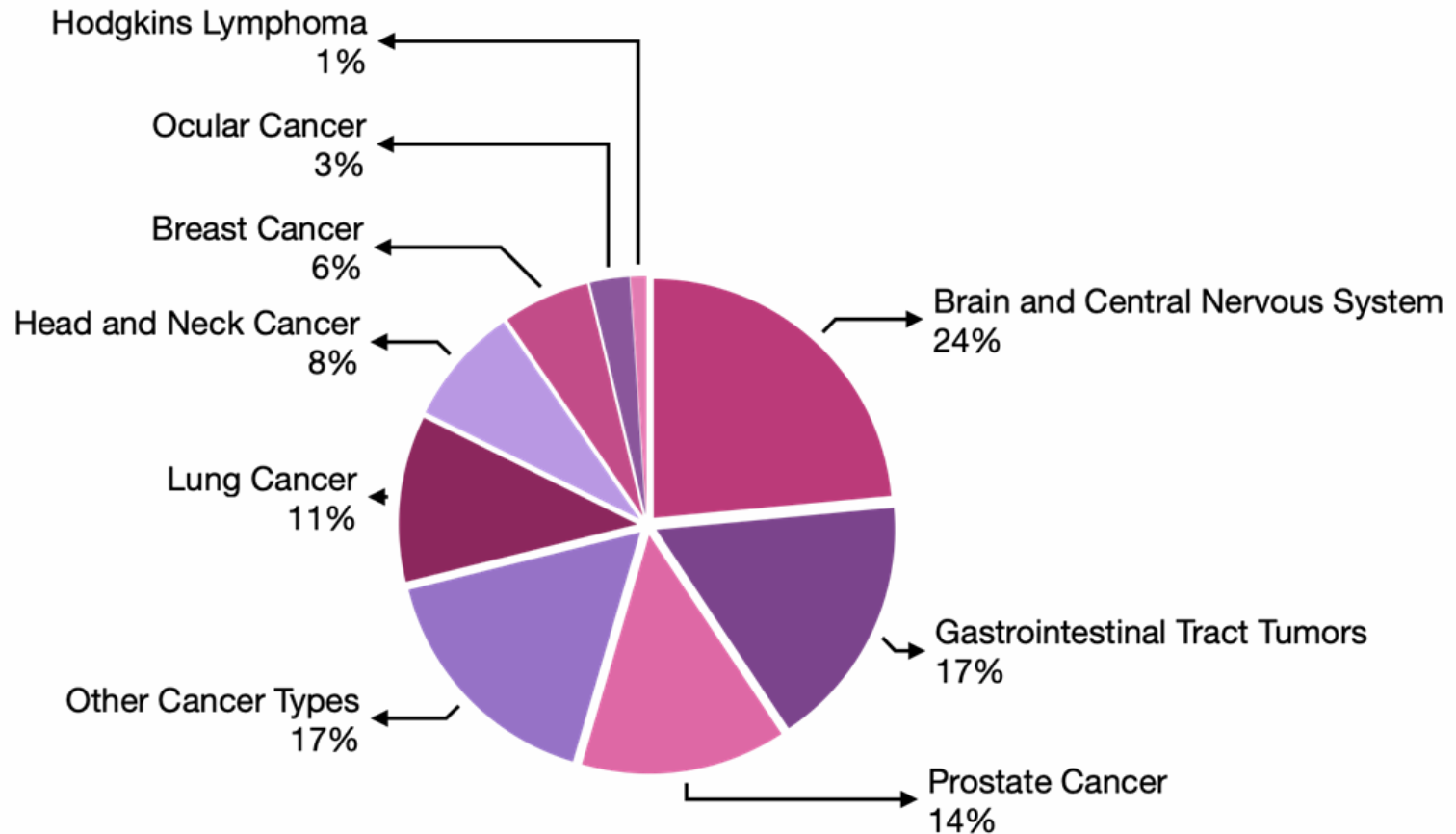


Fig. 6. Percent of clinical trials for different cancer types.



Refence ClinicalTrials.gov ID	Title Status of the trial	Objective Number of participants	Intervention Radiation type	Serious AE Other AE (%)	Authors' Comments
Nantavithya et al. (2018) (NCT01511081)	SBRT vs. SBPT for high-risk early-stage NSCLC Terminated (low accrual)	Phase 2 Randomized to compare SBRT vs. SBPT for side effects, quality of life, cancer control 21	SBRT SBPT	No serious AE with SBRT, metastatic squamous carcinoma of the lung with SBPT Other AE none	No meaningful comparison could be made
Laio et al. (2019) (NCT00915005)	Randomized trial of image-guided adaptive conformal photon vs. PSPT with chemotherapy for NSCLC Completed	Phase 2 Randomized trial to study PSPT vs. IMRT reduces the risk of treatment related pneumonities or tumor recurrence 275	PSPT Photon therapy Paclitaxel Carboplatin	Serious AEs were sig. higher in the PSPT group (38.6%) vs. IMRT (30.4%) p<0.01	PSPT did not provide any benefit in normal tissue toxicity over photon therapy
Lin et al. 2020 (NCT01512589)	Randomized trial of proton beam therapy vs. IMRT for esophageal cancer	Phase 2 Randomized trial to compare PT to IMRT in combination with chemotherapy 180	PT IMRT Fluorouracil Capecitabine Taxane Carboplatin Oxaliplatin	Numerically fewer cardiopulmonary toxicities and post- operative complications in the PT arm	Results are promising, a larger multi- institutional trial is needed

AE – adverse events, NSCLC – non-small cell lung cancer, IMRT – intensity modulate radiotherapy, PT – proton therapy, PSPT – pencil scanning proton therapy, SBPT – stereotactic body proton therapy, SBRT – stereotactic body radiotherapy

Summary of randomized clinical trials that compared proton therapy vs. photon therapy.



Box 1. Key issues that contribute to normal tissue injury with proton therapy

Key Issues	Contributing Factors
Uncertainties in the depth of penetration of beams	<ul style="list-style-type: none">• Differences in tissue compositions• Target movement due to breathing motion, changes in bowel and bladder filling as well as other normal tissue changes
RBE: LET, dose, fraction, and tissue type	<ul style="list-style-type: none">• Contentious use of a fixed RBE value assigned to protons may overdose normal tissue and underdose tumors• Strong impact of LET on normal tissue injury is not being included in the RBE definition• Not using variable RBE values limits personalization of RT
Delineation of the target volume	<ul style="list-style-type: none">• Difficulty in achieving adequate tumor delineation due to want of high-quality diagnostic imaging i.e., CT-MRI or PET-MRI
Target Motion	<ul style="list-style-type: none">• Changes in tumor location during treatment fractions caused by:<ul style="list-style-type: none">○ discrepancies between day-to-day patient set up procedures○ breathing○ peristaltic movement○ bowel movement○ heart beats○ organ filling

Abbreviations: CT-MRI, computed tomography-magnetic resonance imaging; LET, linear energy transfer; PET-MRI, positron emission tomography-magnetic resonance imaging; RBE, relative biological effectiveness; RT, radiotherapy

- ✓ Differences in treatment plans,
- ✓ Fractionation schemes,
- ✓ Total dose,
- ✓ Patient characteristics,
- ✓ Tumor location,
- ✓ Medications and other agents, surgical history etc.



- ✓ Despite technological advances in RT patients still experience adverse effects. Irradiation of normal tissue is almost unavoidable. Normal tissue is an important determinant of the outcome.
- ✓ While a steady increase in PT literature is observed, mostly non-randomized early phase clinical trials with a relatively small number of patients are enrolled. Retrospective analysis and single arm studies will likely favor PT.

Proton therapy with favourable dose delivery can improve treatment outcomes vs. benefit in normal tissue toxicity

Dosimetric benefit does not ensure PT has more favorable clinical outcomes but does form a basis for comparative prospective trials.

To fully benefit from PT and justification of costs, comparative and prospective large-scale well-designed clinical trials are necessary incorporating suitable biological determinants (panels before, during or after) to optimizing treatment process.

Proton Therapy: the Problem? Does it make any sense to spend over \$100 million on a proton facility, with the aim to reduce doses to normal tissues and then to bathe the patient with a total body dose of neutrons”, Hall, Technol in Ca Res Treat 2007;6:31-34.

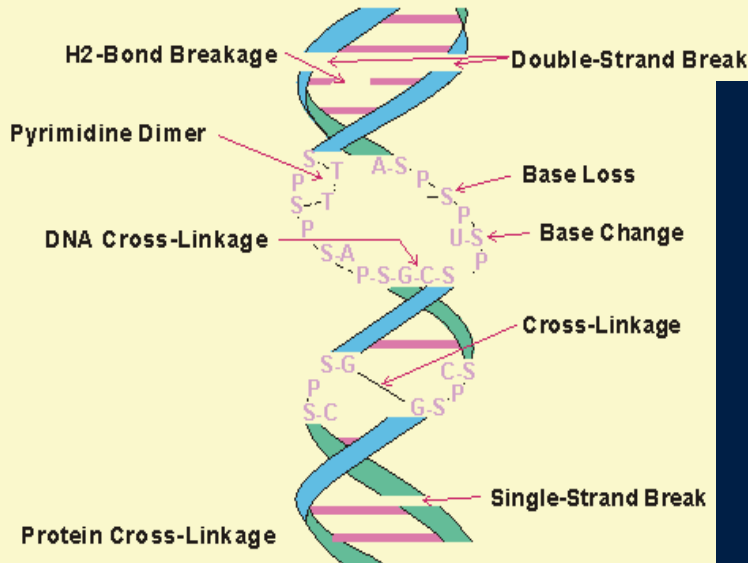


Experimental studies on the biological effects of proton irradiation are relatively sparse and focus mostly on different cancer cells response and RBE.

There is an unmet need to recognize the possible differences in the mechanisms of radiation injury and the normal tissue toxicities with different radiation types.



RADIATION DAMAGE TO DNA



The DNA is the critical target to induce lethal effects as a result of radiation exposure.

Radiation can induce damage either by direct action or indirect action (free radicals).

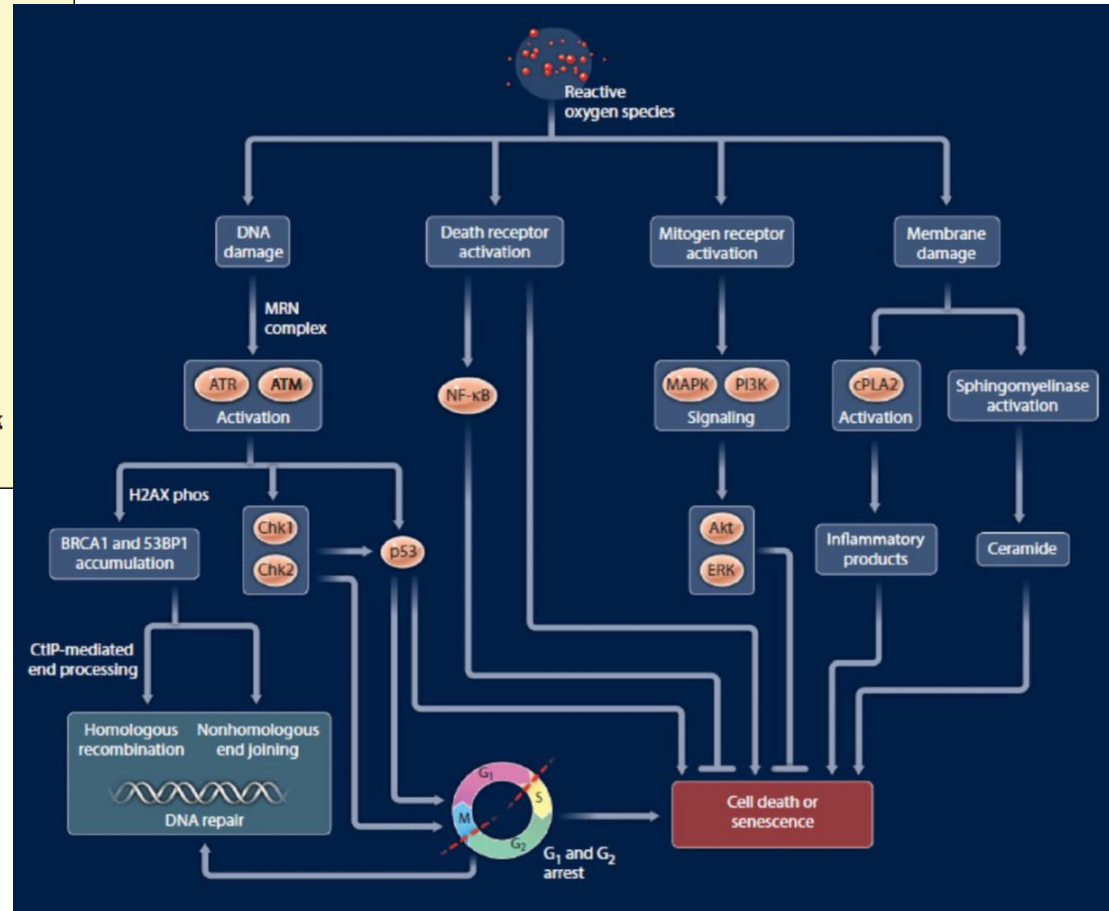


Fig. 7. Various cellular targets and responses that occur after radiation exposure.



A modulation of normal tissue response can be expected in all dose ranges



Circulating lymphocytes represent normal tissue and traffic throughout the body including irradiated tumor volume.

Following a standard regime of thirty fractions of 2Gy, 98.8% of the blood pool has been exposed to more than 0.5 Gy.

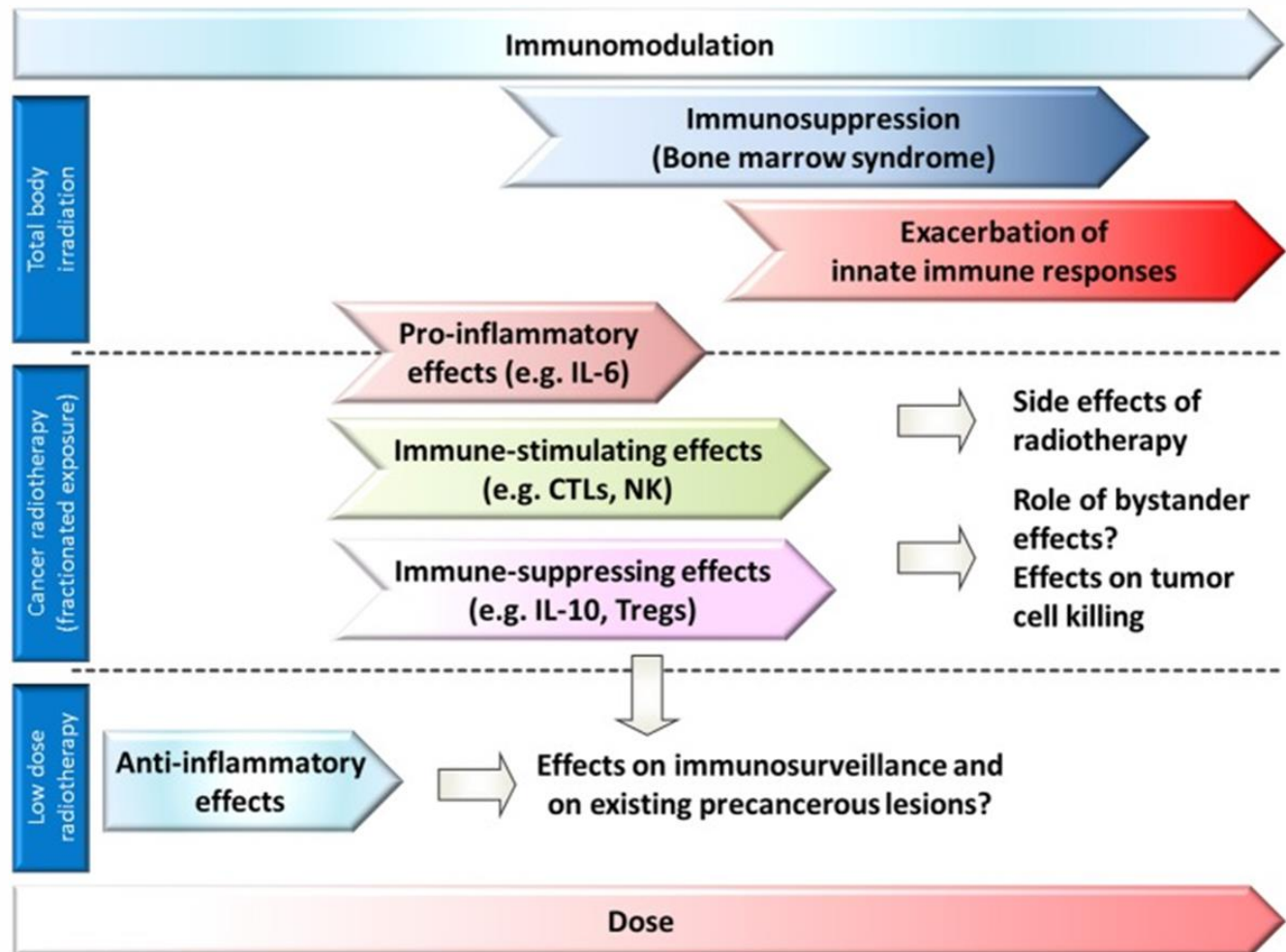
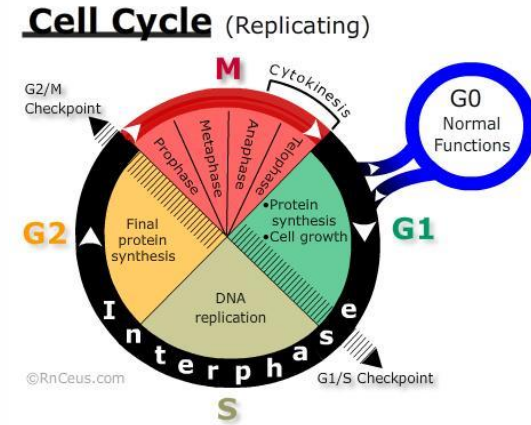


Fig. 8. Role of immune response in different dose ranges after WBE and PBE.



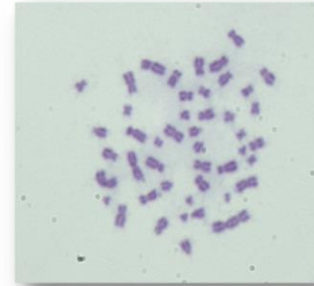
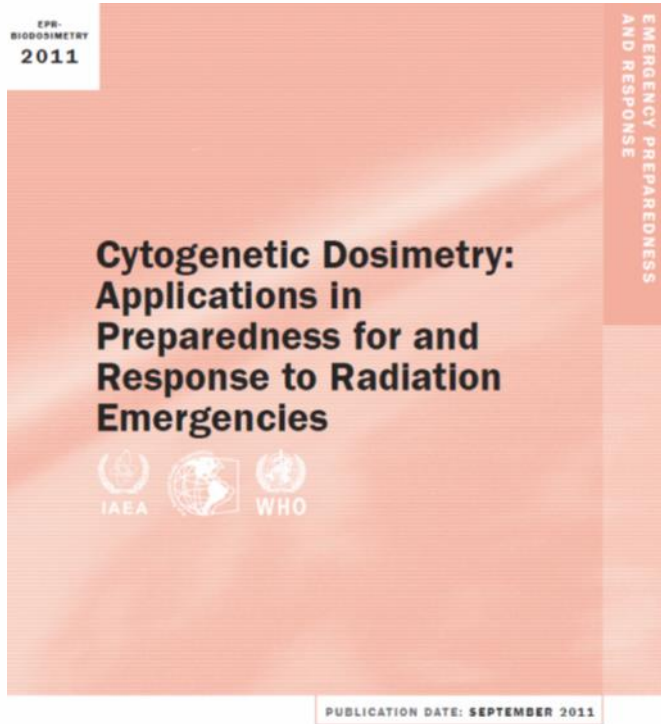
- Predominantly in a resting G0 phase, consist of T and B cells.
- Synchronous and homogeneous cell population.
- Circulate throughout the body and can be stimulated to undergo mitosis, recirculation time~ 12h, 80%. Even when a small part of the body was irradiated – dosimetry is possible.
- Involved in many key mechanistic roles following exposure to radiation therapy, persistence of DNA damage 3.5 years.
- Used to interrogate radiation injury to normal tissue during tumors irradiation (critical targets for radiotherapy and immunotherapy). Blood taking is not-invasive.



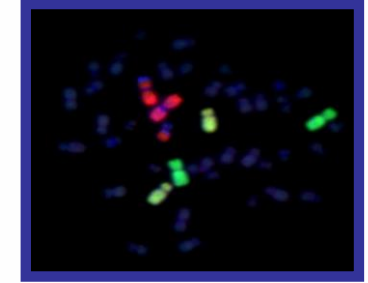
HPBL is a well-accepted model to study the effects of radiation on normal tissue.



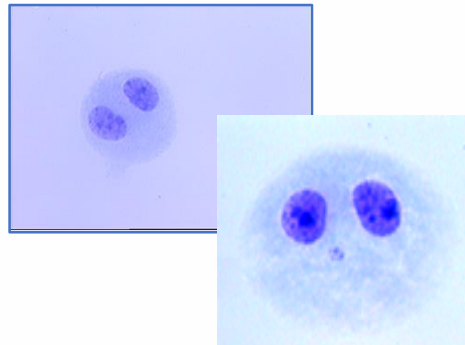
DSBs are critical lesions and their misrepair or non-repair are involved in the formation of chromosome aberrations.



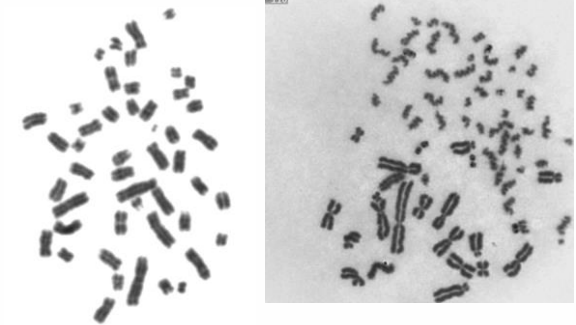
Dicentric analysis (DA)



Translocation analysis (FISH)



The cytokinesis-block micronucleus (CBMN)

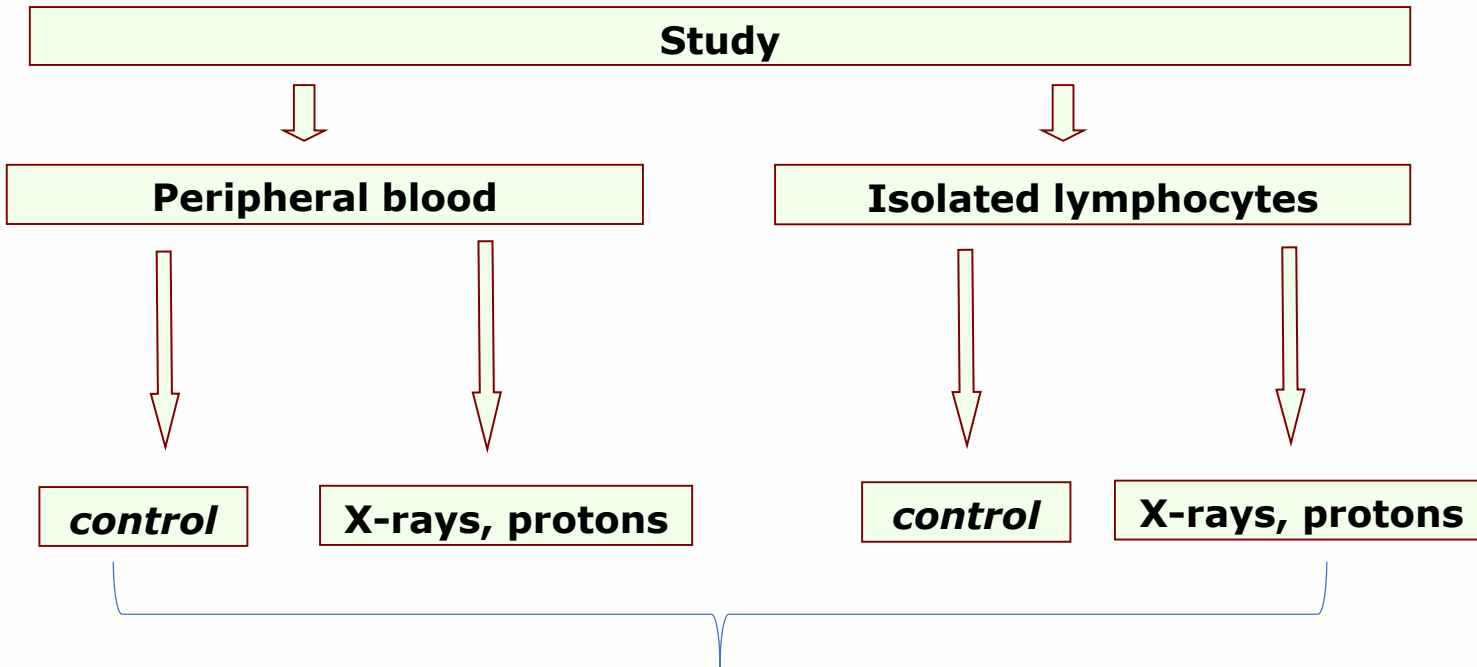


Premature chromosome condensation (PCC)



Time-consuming, expensive, up to 4.0 Gy, protocols...

Every laboratory must optimize protocols and obtain own calibration curves.



- To prepare phantom, laboratory and optimize methods for low and high radiation dose assessment
- To study the response of normal tissue after protons vs. photons (DNA damage, cell death, cell cycle)
- To propose mechanisms of molecular and cellular injury by protons in HPBLs.

Approval from the human bioethical committee of the Regional Medical Board (No. 124/KBL/OIL/2013) and (184/KBL/OIL/2020).

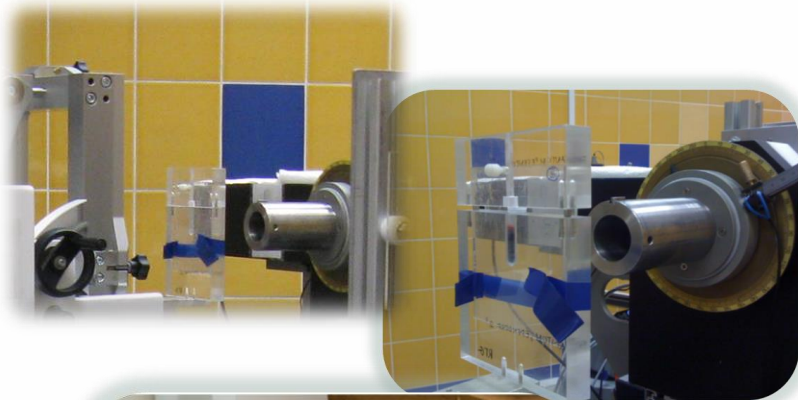


Proton radiobiology

Response of human lymphocytes to proton radiation of 60 MeV compared to 250 kV X-rays by the cytokinesis-block micronucleus assay

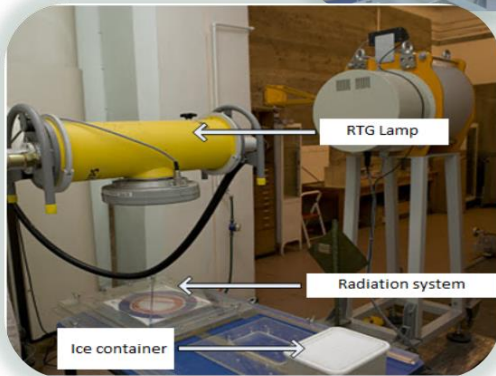


Justyna Miszczyk^{a,*}, Kamila Rawojć^{a,b}, Agnieszka Panek^a, Jan Swakoń^c, Pataje G. Prasanna^d, Marzena Rydygier^c



Whole blood or isolated lymphocytes
(10 donors, ♀ 42.0±4.5, ♂ 44.0±5.6
were irradiated in the dose range from 0.3 – 20.0 Gy
(0.3, 0.5, 0.75, 1.0, 1.5, 2.0, 2.5, 3.0, 4.0)
(8.00, 13.64, 15.00, 20.00)

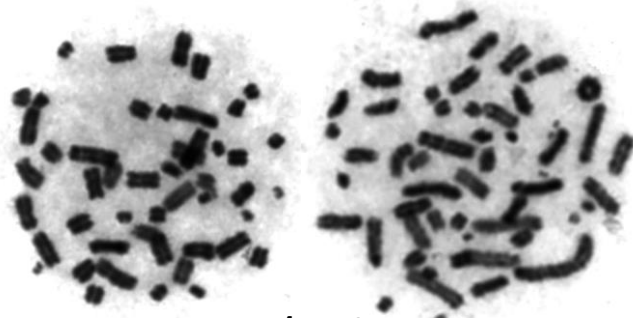
Cells were irradiated with a 60 MeV proton beam in 2-cm Eppendorf vials. A specially designed PMMA - Poly(methyl methacrylate) phantom was placed at the irradiation setup isocentre (in the middle of SOBP) and in the centre of the flat beam. Average dose rate 0.075 Gy/s.



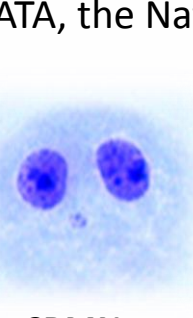
The radiation was delivered at a dose rate of 0.15 Gy/s by a Philips X-ray machine (MCN 323 model, Philips) at 250 kV. The vials were placed in a polyethylene box; radiation field-size was 20x20 cm², and the source to surface distance was 34.8 cm.



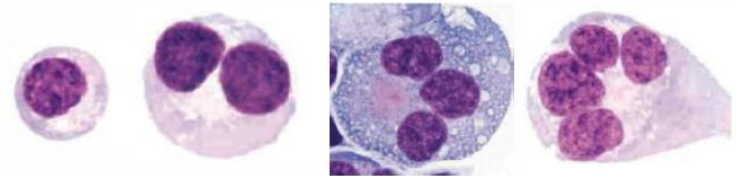
“Development and optimization of PCC (*Premature Chromosome Condensation*) method for the purposes of proton radiotherapy at the Cyclotron Center Bronowice”, SONATA, the National Science Centre.



PCC, G2/M phase



CBMN



Laboratory of spectroscopic imaging for radiobiology, treatment and research of complex systems

Laboratory of spectroscopic imaging for radiobiology, treatment and research of complex systems

Laboratory of spectroscopic imaging for radiobiology, treatment and research of complex systems

Goals of the project:

Creation of a modern region-wide laboratory, capable of carrying out unique research on the impact of ionizing radiation on living organisms and providing research facilities for the Bronowice Cyclotron Centre.

Conducting research, using physical methods, for the purpose of eradicating cancer and other pathologies. This includes spectroscopic imaging of cells and tissues (examining the internal structure of cells, the cytoskeleton organization, its mechanical and biochemical properties) as well as research at the molecular level.

Carrying out fundamental research in the field of structural and vibrational analysis of condensed matter.

The Integrated Raman-AFM system capable of performing Raman microspectroscopy, atomic force microscopy (AFM) and Raman nanospectroscopy enhanced on the AFM tip (TERS).

The NanoIR system - integrated IR-AFM layout - infrared microscope with nanometer resolution achieved by enhancing signal on the AFM tip.

Vacuum FTIR spectrometer (VERTEX 70 FT-IR spectrometer with a microscope HYPERION 3000) with a microscope, bolometer and FPA detector. Operation range: from near to far infrared. Purpose: taking chemical characteristics of the sample, including studies of radiation damage to cells and tissues, and interaction with chemotherapeutic agents.

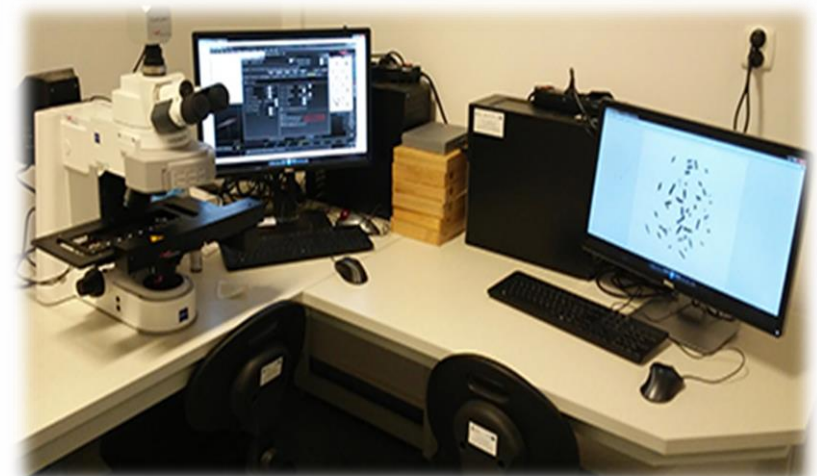
UV-VIS Spectrometer plate reader - for the laboratory accreditation requirements, allowing the use of multiple absorption, fluorescence and luminescence markers.

Atomic force microscope integrated with a fluorescent microscope.

System for automatic analysis of chromosome aberrations, Upgrading our existing system (Zeiss microscope Axiolmager Z2) for karyotype determination. The extension includes capability of automatic metaphase searching and building up a test stand of GTG and FISH analysis for accreditation requirements.

Independent fast imaging IR microscope (Nicolet IN10 MX) - for spectral imaging and spot measurements, equipped with three detectors and motorized microscope table. Capable of recording the spectrum in both transmission and reflection modes, as well as of taking classical microscope images of the sample.

The RT-PCR system for real-time testing of DNA sequence, fulfilling accreditation requirements. Used in the research involving: gene expression, single nucleotide polymorphism (SNP), dysfunctional copy number of the gene (CNVs), pRNA profiles, detection of pathogens.



High-performance slide scanning Metafer platform (Axiolmager.Z2) microscope with specimen holder for 8 slides (76x26 mm) with two independent Ikaros software modules for automatic cells detection and analysis.

The Henryk Niewodniczański Institute of Nuclear Physics, Polish Academy of Sciences in Krakow



Multimodal approach for ionizing radiation damage investigation, Kwiatek W. et al.

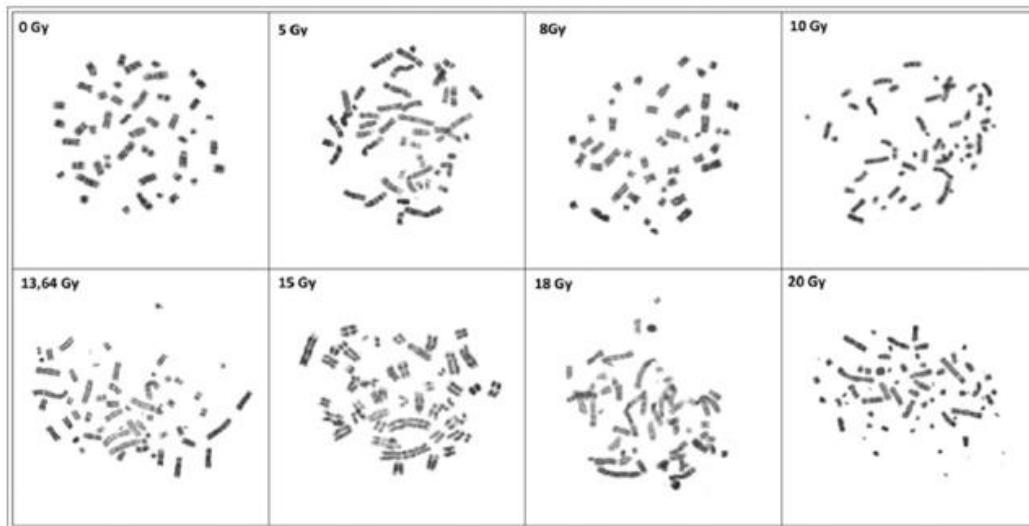
<https://los.ifj.edu.pl/en/index.html>

HUMN project: detailed description of the scoring criteria..., Fenech M. et al.



Evaluation of the premature chromosome condensation scoring protocol after proton and X-ray irradiation of human peripheral blood lymphocytes at high doses range

K. Rawojć, J. Miszczyk, A. Mozdzeń, J. Swakoń & A. Sowa-Staszczak



PCC micrographs in G2/M phase post proton irradiation

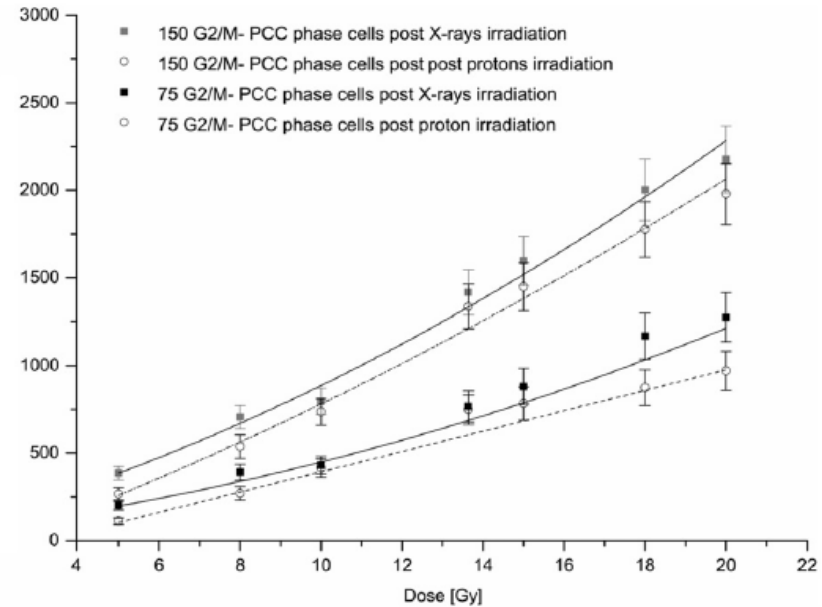


Fig. 9. Comparison between 150 vs. 75 G2/M cells scoring modes for all scorers and both types of radiation.

Similar distribution trends for both scoring modes were observed.



Effects of culturing technique on human peripheral blood lymphocytes response to proton and X-ray radiation

Justyna Miszczyk & Kamila Rawojć

The isolation process did not significantly influence the cell proliferation ability after irradiation.

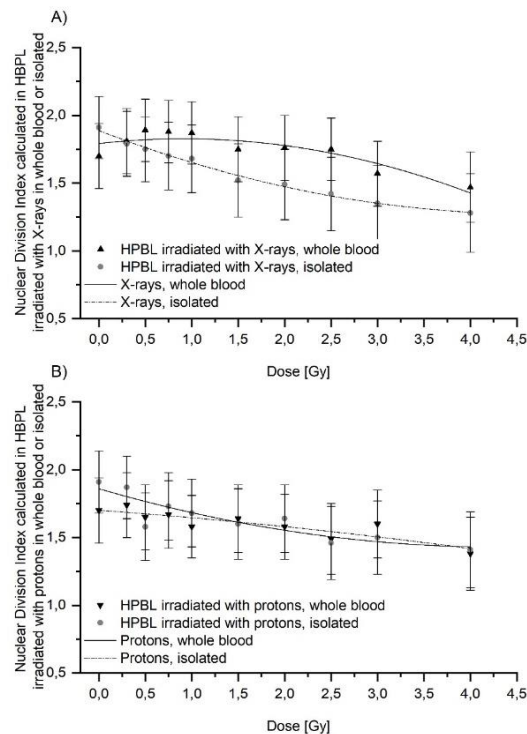


Fig. 10. Proliferation index (NDI) in irradiated cells with X-rays and protons, blood vs. isolated cells.

Significant differences in DNA damage are evident and affected by the radiation type.

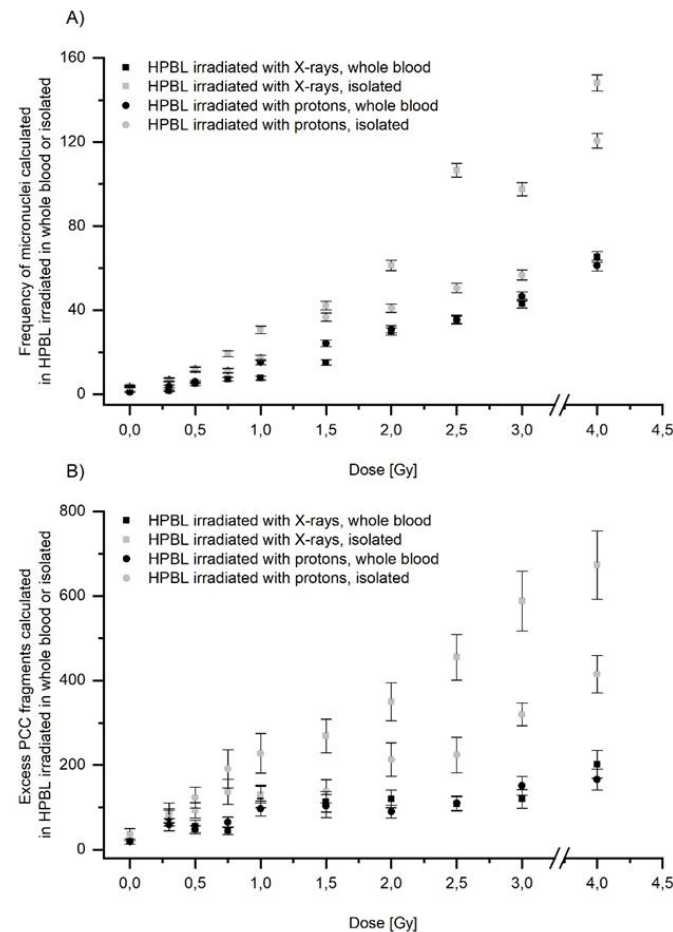


Fig. 11. DNA damage in irradiated cells with protons and X-rays, cultured in two different techniques.



Similar trends in estimated biomarkers between different treatment conditions were observed.

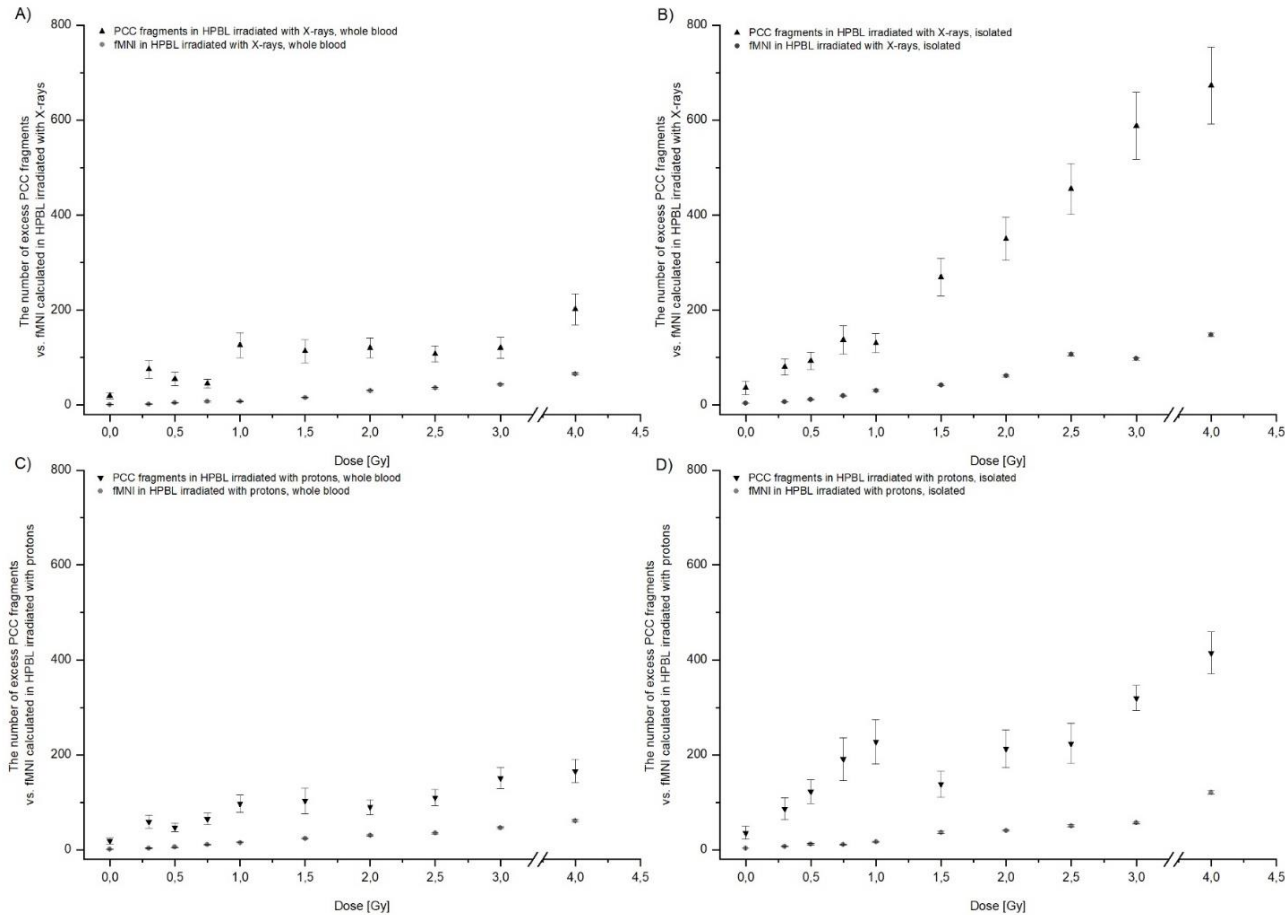
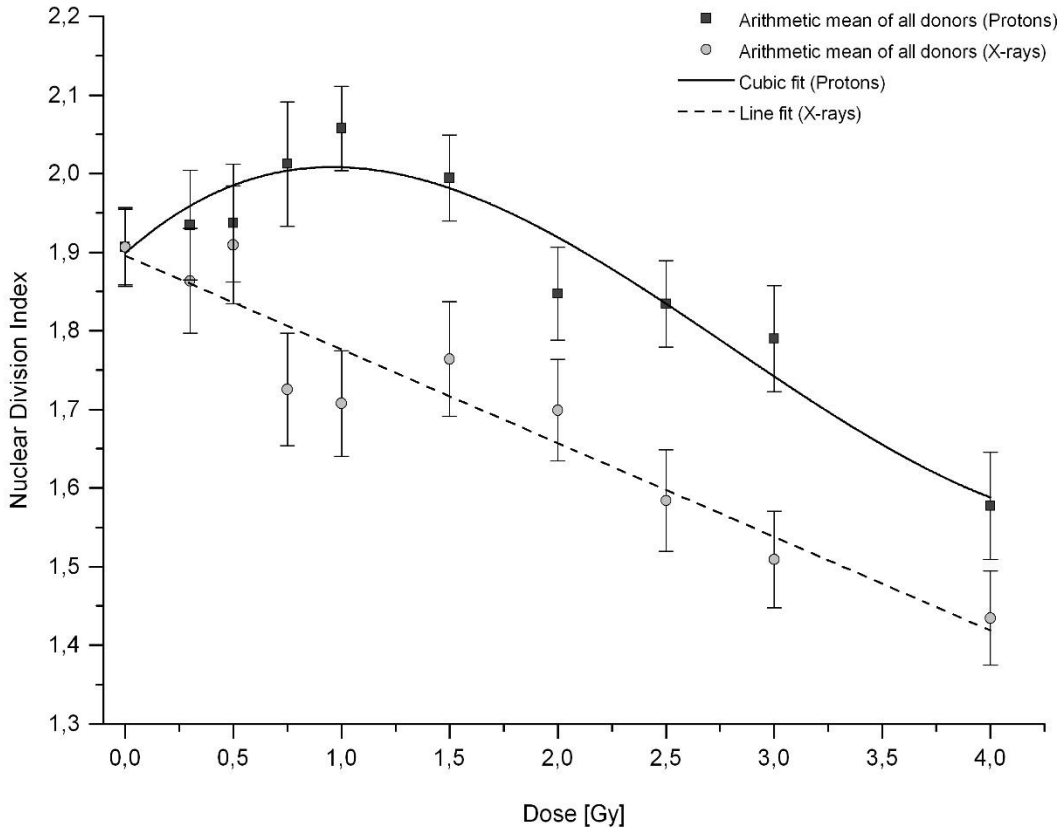


Fig. 12. Comparison of biomarkers in isolated or whole blood cultured lymphocytes post X-ray and proton irradiation.



Compared to X-rays, cellular proliferation after irradiation with protons was significantly higher in HPBL for all doses within the range of 0.75-4.0 Gy.

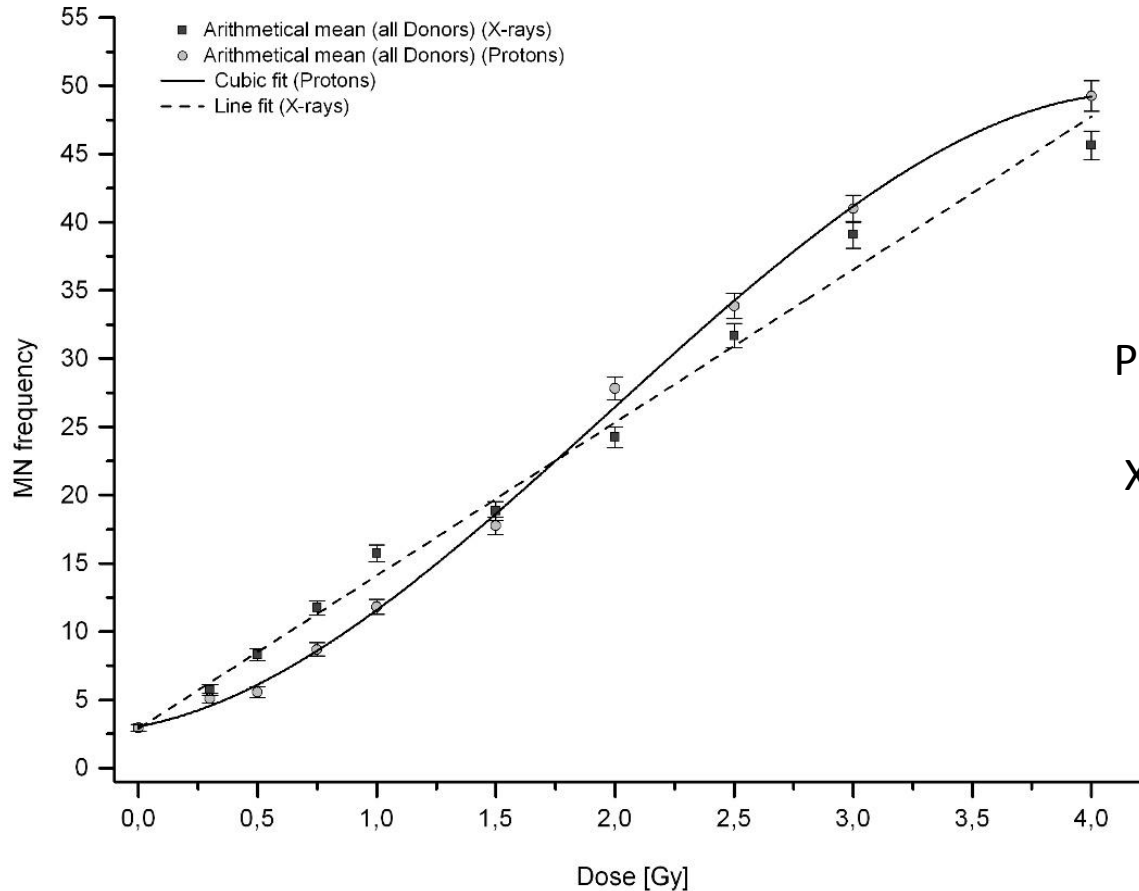
At dose above 1.0 Gy for both radiation types a gradual and progressive decline was seen.

Fig. 13. Dose-effect relationship for NDI in HPBLs following irradiation with protons and X-rays.

Protons and X-rays influence HPBL cellular proliferation to different degrees.



Most likely, protons and photons induce DNA damage in HPBL by different mechanisms.



Protons were significantly less effective as compared to X-rays at doses below 1.5 Gy.

Fig. 14. Dose-response curves for DNA damage for both radiation types.



Extent of difference varies among donors without correlation with radiation type.

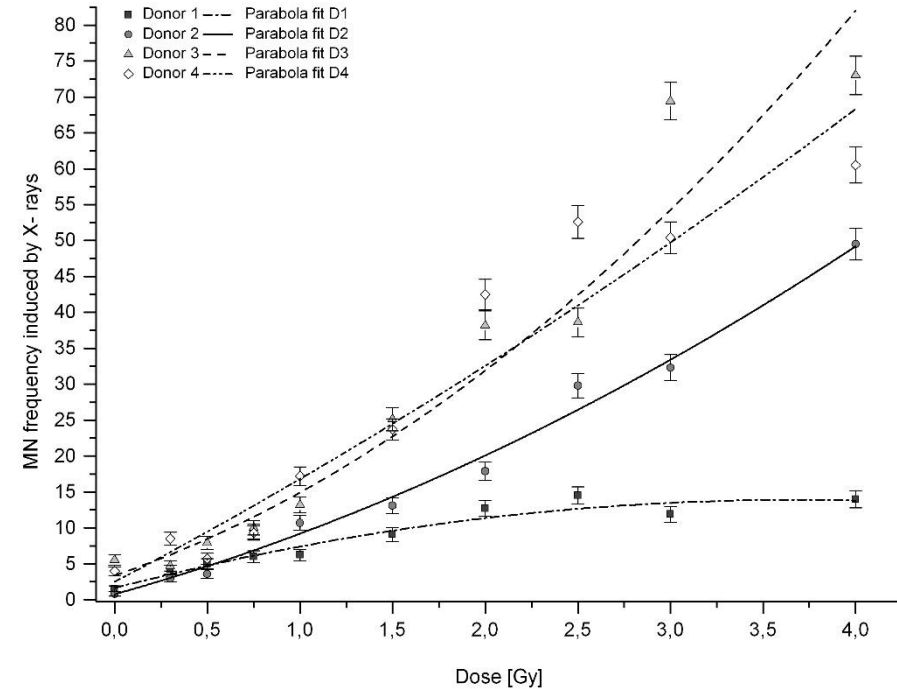
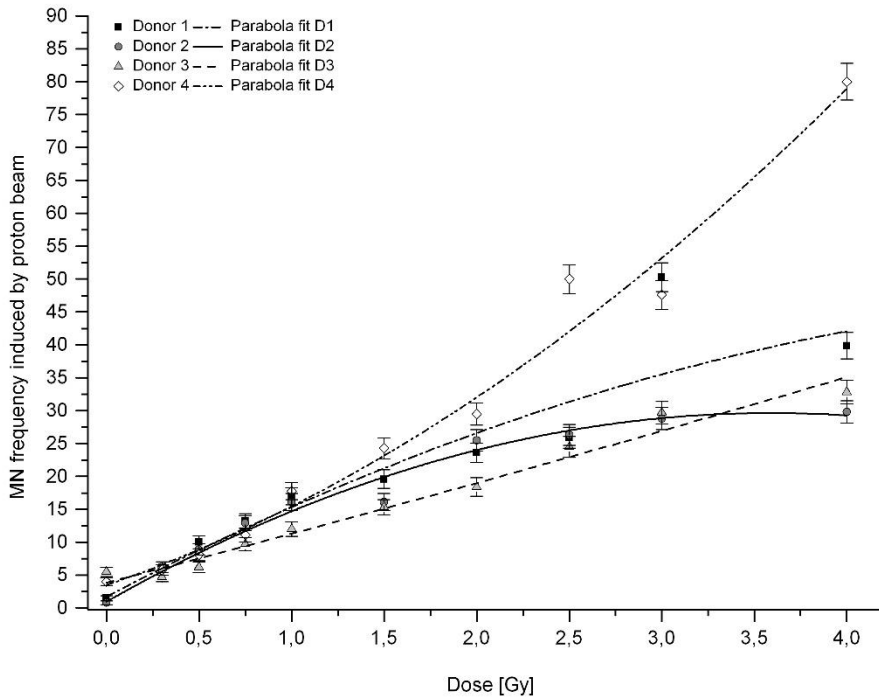


Fig. 15. The dose-dependent relationship between DNA damage for the proton beam and X-rays among individuals.

At lower proton doses curves overlapped with each other, differences were more pronounced at doses above 1.0 Gy.



Distribution of DNA damage following irradiation with protons and photons is different.

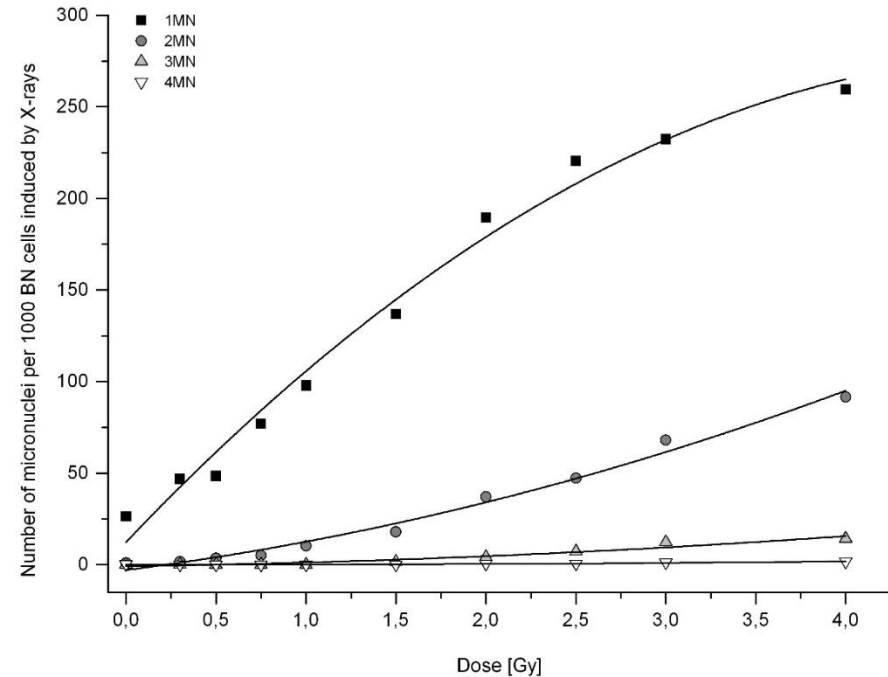
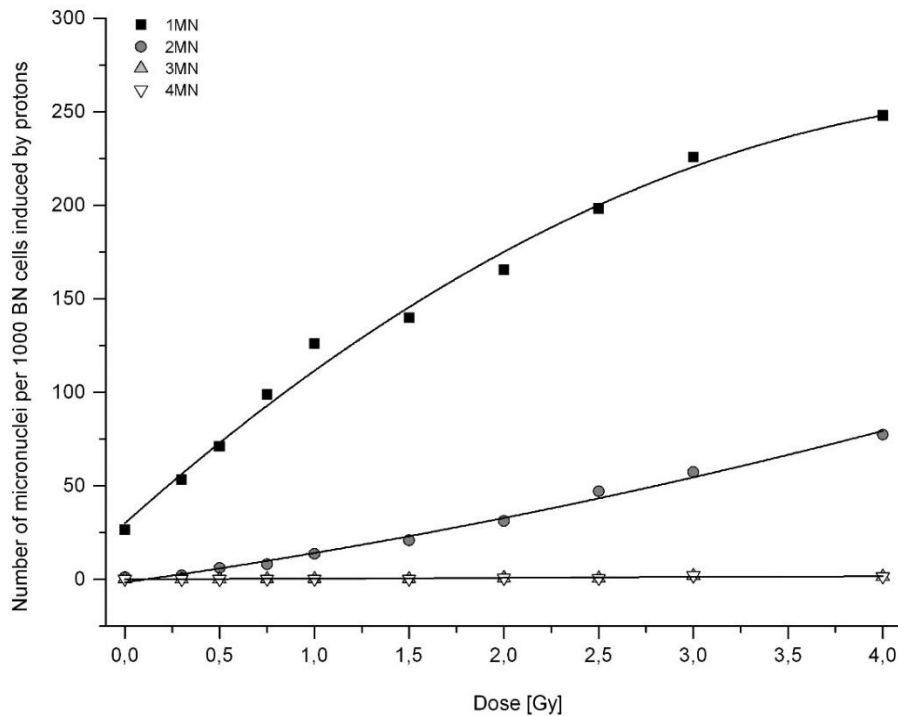


Fig. 16. The dose-response relationships for the protons and X-rays and various number of micronuclei (1, 2, 3 and 4) in 1000 BN cells as an average for all donors.

Locally multiply damaged sites or clustered DNA damage?



This article belongs to the Special Issue

Applied Physics in Cancer Cells

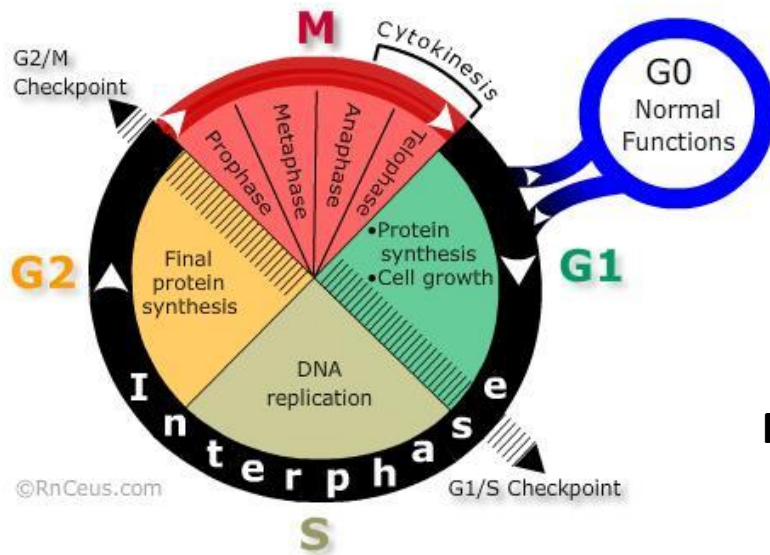


Article

Investigation of DNA Damage and Cell-Cycle Distribution in Human Peripheral Blood Lymphocytes under Exposure to High Doses of Proton Radiotherapy

Justyna Miszczyk

Cell Cycle (Replicating)



Cell death

Mitotic delay

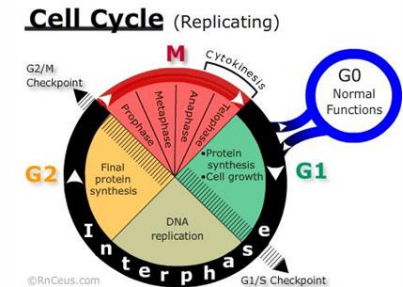
The effects and mechanisms of a single high-dose delivery remain unclear.

Pre-clinical studies have suggested that RT, especially higher doses of 20-25 Gy can substantially stimulate anti-tumor T-cell immunity and increase the T-cell response to help control tumor growth.



X-rays	G1	S	G2	M	A	Nucleated
8.00	0.05	1.90	4.45	0.05	0.25	93.30
13.64	0.05	1.60	2.20	0.00	0.00	96.15
15.00	0.00	1.50	2.00	0.00	0.00	96.50
20.00	0.00	1.25	1.45	0.00	0.00	97.30

Protons [Gy]	G1	S	G2	M	A	Nucleated
8.00	0.00	1.65	2.80	0.05	0.70	94.80
13.64	0.00	1.85	1.35	0.00	0.20	96.60
15.00	0.05	1.40	1.10	0.00	0.30	97.15
20.00	0.10	1.25	0.90	0.00	0.20	97.55



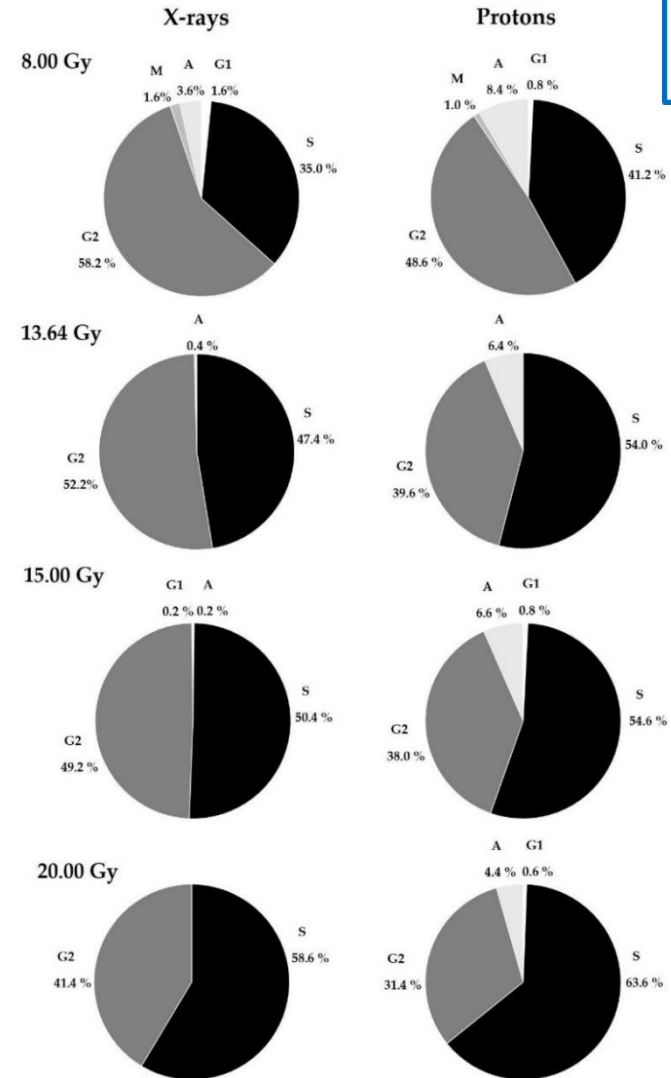
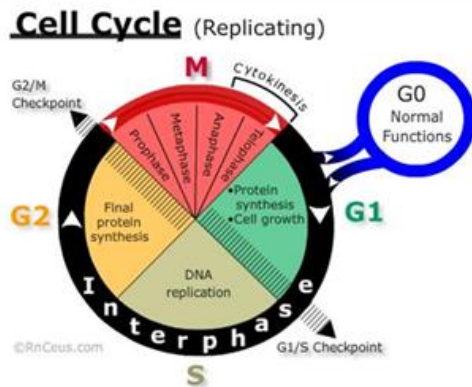
Percentages of G1, S, G2, M, A phases and nucleated cells at each dose of X-rays or protons.

As the dose increases, the number of nucleated cells for both studied types of radiation increases to comperable values.



The largest proportion of cells were
in the S and G2 phases.

For both types of radiation, as the dose increased, the number of S-phase cells also increased and was higher for each dose after proton radiotherapy.



Percentages of cells in the G1, S, G2, M, A phases at each dose after X-rays or protons.



High doses of protons induce DNA damage in the G2/M differently than X-rays.

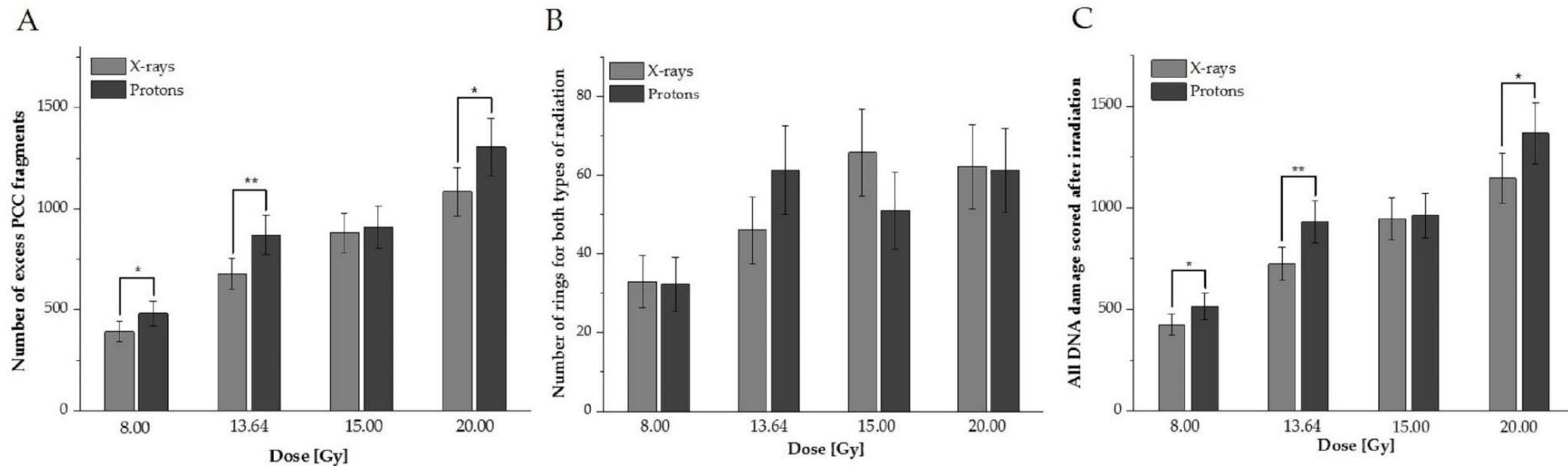


Fig. 17. Average values of DNA damage for HPBLs irradiated with high doses.

For all doses except 15.00 Gy, a higher DNA damage value after proton therapy was observed.



The distribution of DNA damage following high doses of irradiation with protons vs. photons differs between donors, types of radiation, and doses.

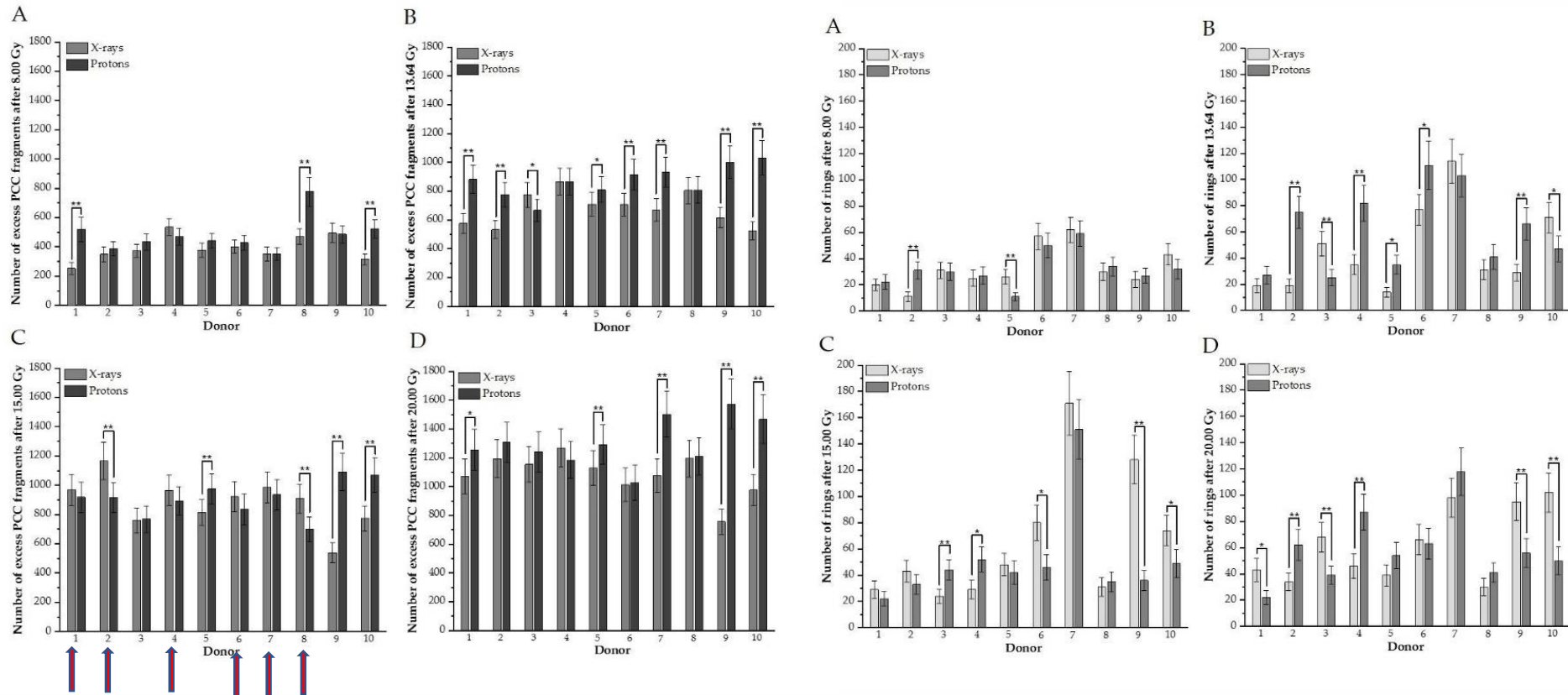


Fig. 18. An individual number of DNA damage for donors irradiated with different doses of protons vs. X-rays.



More scattered distribution for protons vs. X-rays was observed.

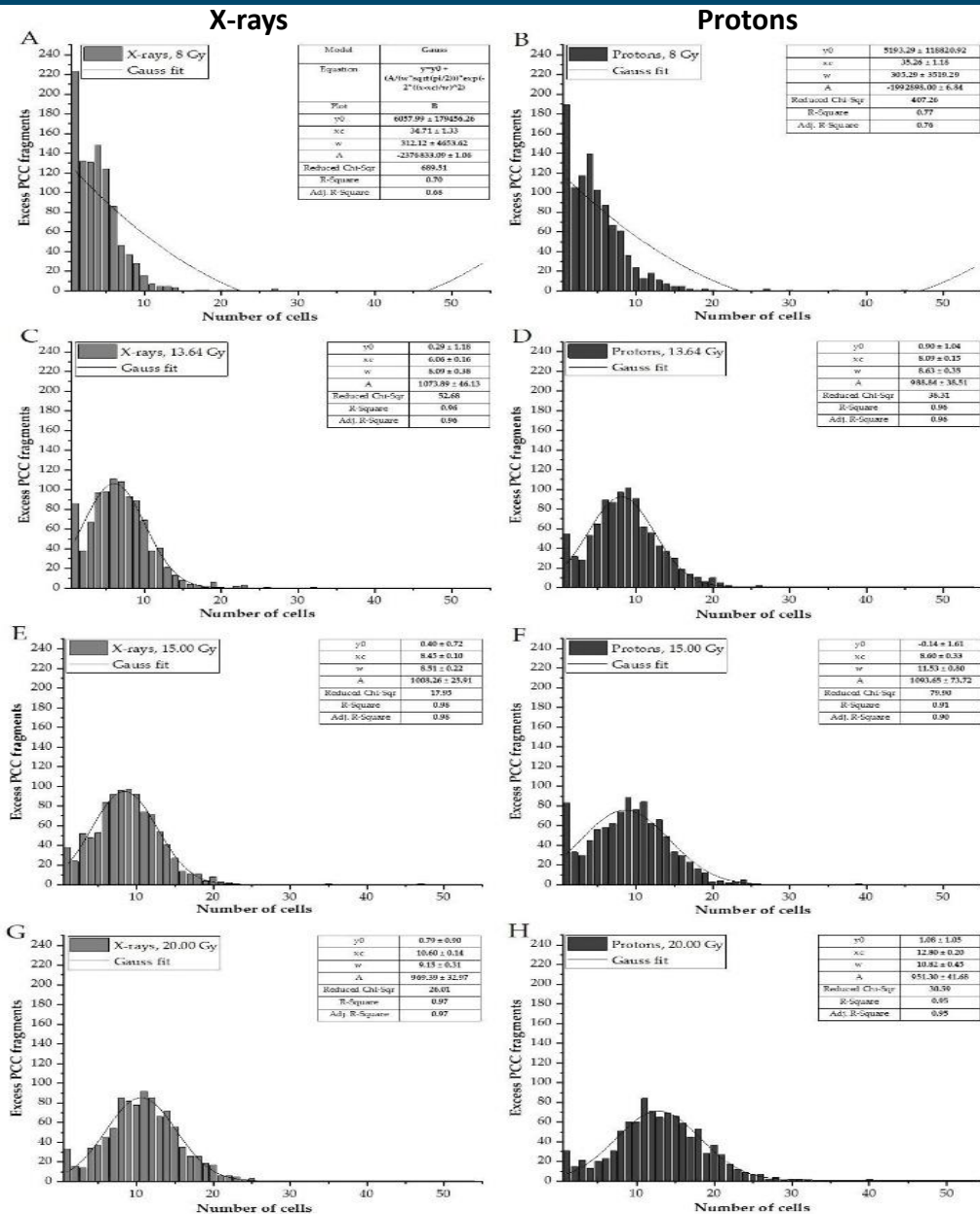
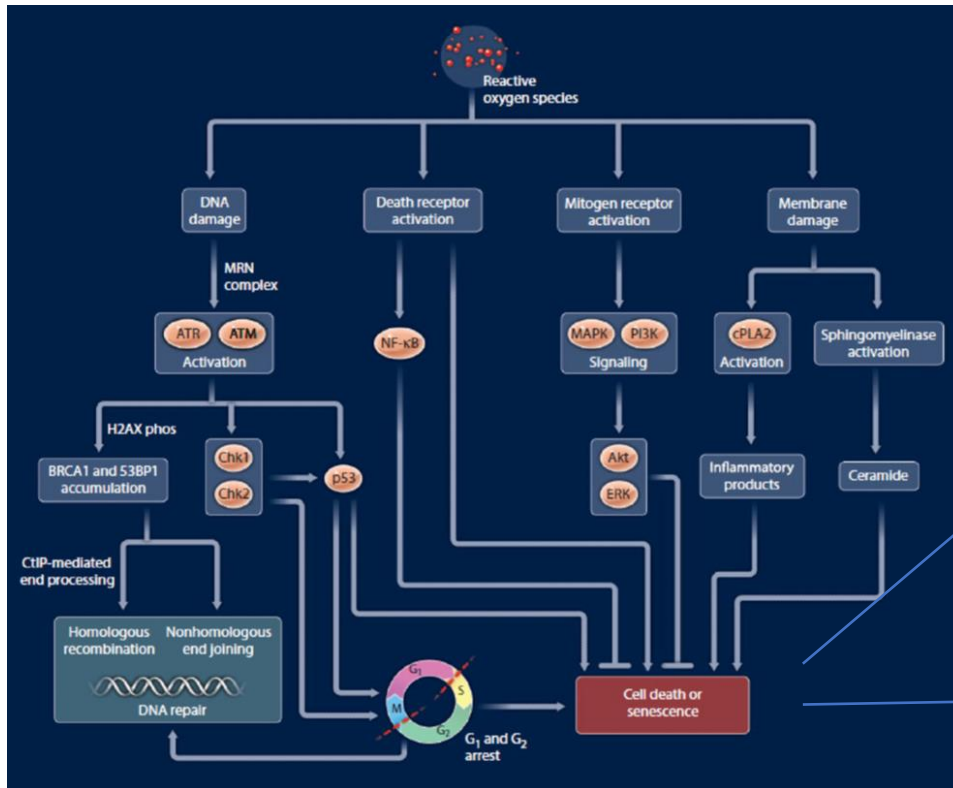


Fig. 19. Frequency distribution of the number of DNA damage after X-rays or protons.



Apoptosis and necrosis are controlled by different molecular pathways.



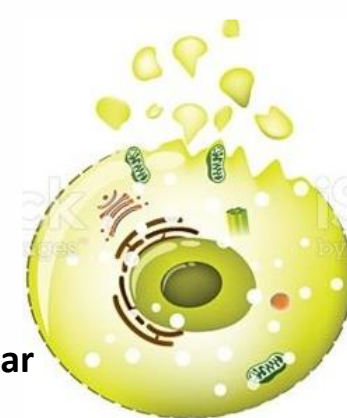
Cell membrane remains intact,
cell breaks into apoptotic bodies.
No inflammation.

**Apoptosis
(programmed
cell death)**



Necrosis

Loss of cell membrane
integrity, resulted in an
uncontrolled release of
products into the extracellular
space. **Inflammation.**





Clinical and Translational Radiation Oncology 9 (2018) 23–29

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Clinical and Translational Radiation Oncology

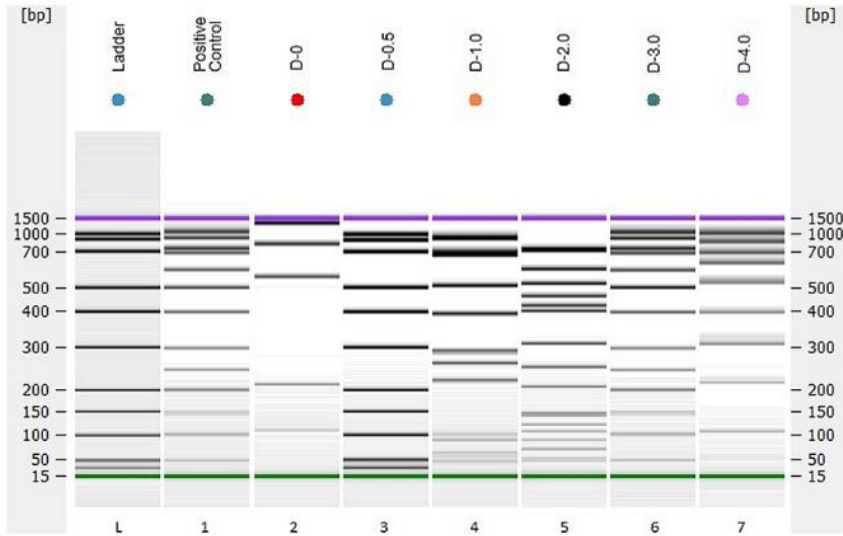
journal homepage: www.elsevier.com/locate/ctro



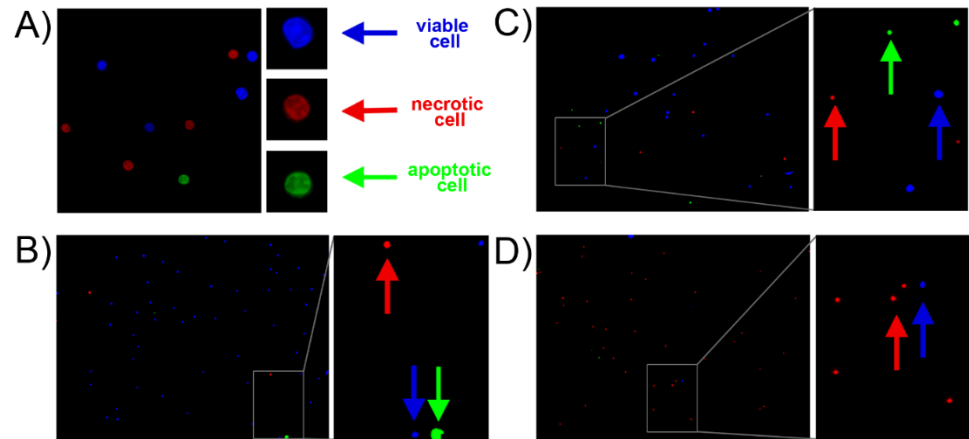
Original Research Article

Do protons and X-rays induce cell-killing in human peripheral blood lymphocytes by different mechanisms?

J. Miszczyk^{a,*}, K. Rawojć^b, A. Panek^a, A. Borkowska^a, P.G.S. Prasanna^c, M.M. Ahmed^c, J. Swakoń^a, A. Gałaś^d



DNA (chip) fragmentation assay confirmed various degrees of DNA fragmentation. Lanes showed apoptosis, as well as smearing indicative of necrosis.



Cells after ex vivo irradiation with 3 Gy protons following staining with Apoptotic, Necrotic and Healthy Cells Kit, (Biotium, Hayward, USA).

Proton irradiation resulted in higher apoptotic activity.

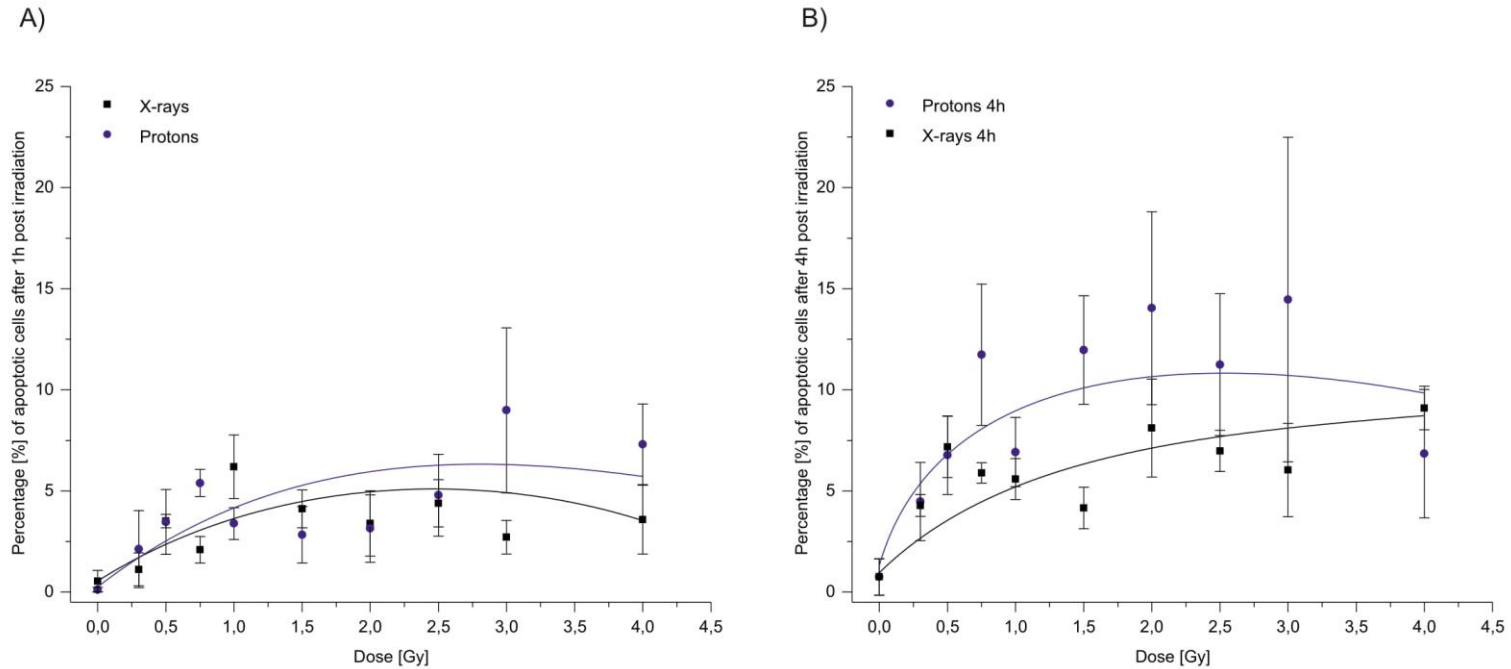


Fig. 20. Percentage of apoptotic cells after irradiation with X-rays vs. protons (1 and 4 h after irradiation).



Original Article

Translational Cancer Research 2018;
7(4):879-889.



Therapeutic proton irradiation results in apoptosis and caspase-3 activation in human peripheral blood lymphocytes

Justyna Miszczyk¹, Kamila Rawojć², Anna Maria Borkowska¹, Agnieszka Panek¹, Jan Swakon¹, Aleksander Gałaś³, Mansoor M. Ahmed⁴, Pataje G. S. Prasanna⁴

Variability between donors and types of radiation is observed.

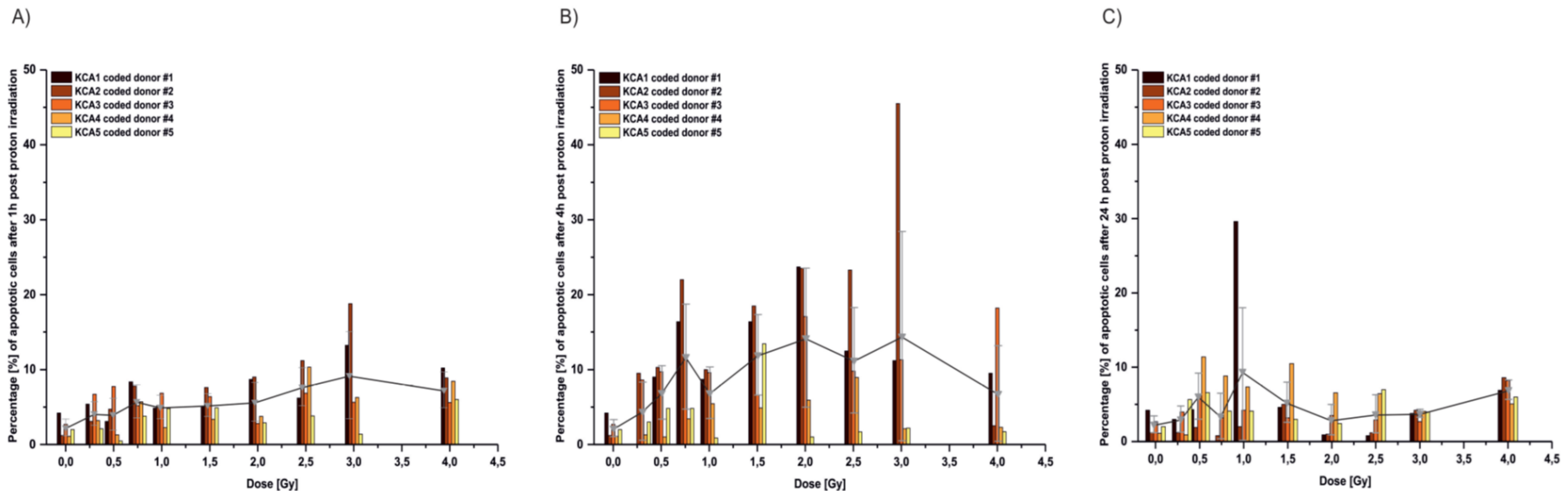
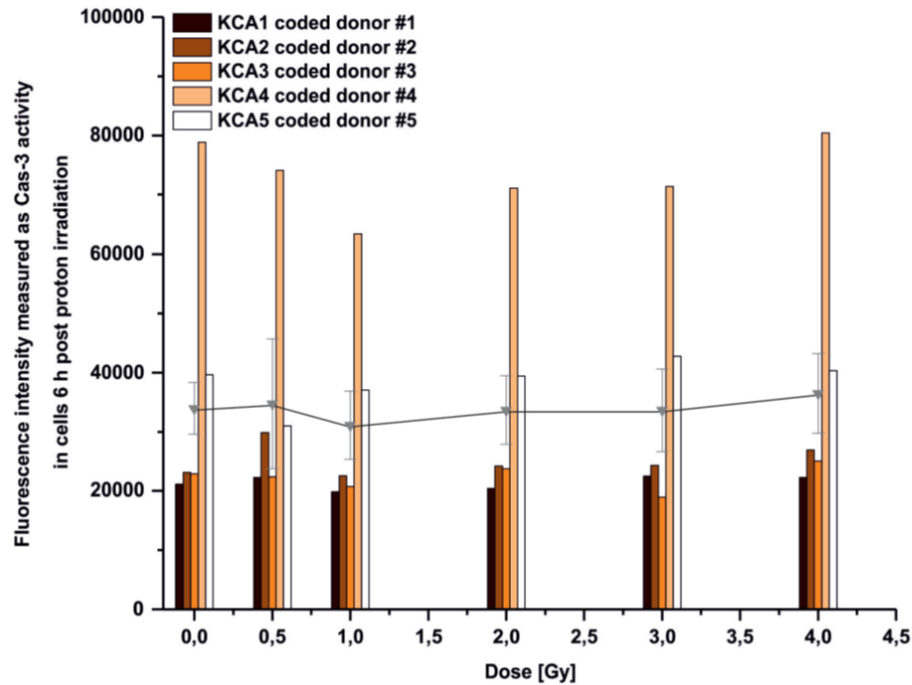


Fig. 21. Apoptotic cells percentage for individuals determined with Annexin V-FITC fluorescent staining in *ex vivo* HPBL model after 1 h, 4 h and 24 h post-proton treatment.

After proton irradiation apoptosis is mediated through caspase-3,
activity is mostly observed at 6 h proton-irradiation, decreased significantly after 24h.

A)



B)

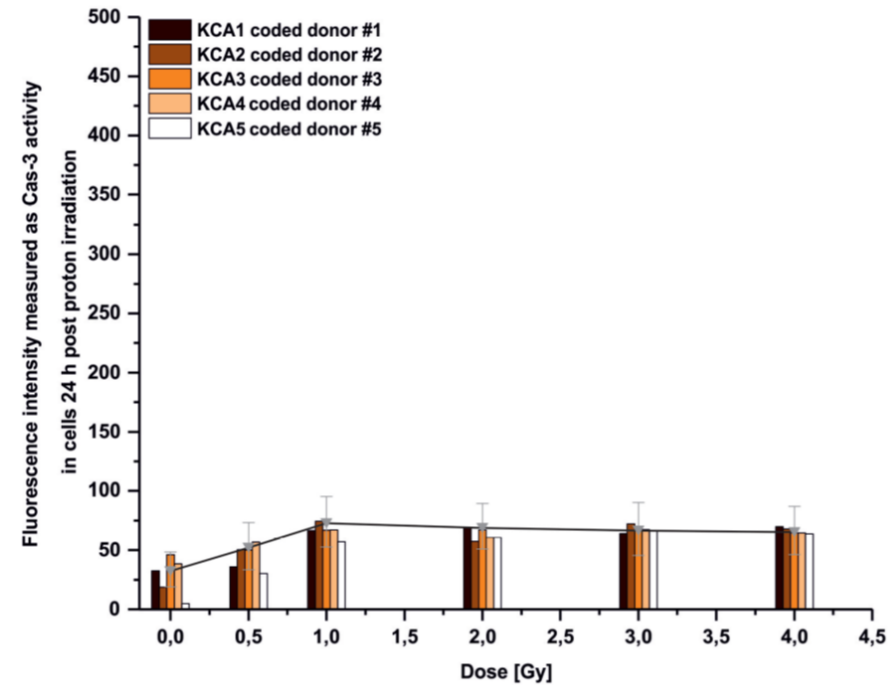


Fig. 22. Differences in caspase-3 activity among donors at 6 and 24 h post proton treatment (HTS assay kit, Biotium).

Protons are more efficient in cell-killing due to their potential to cause necrosis in addition to apoptosis, especially at higher doses!

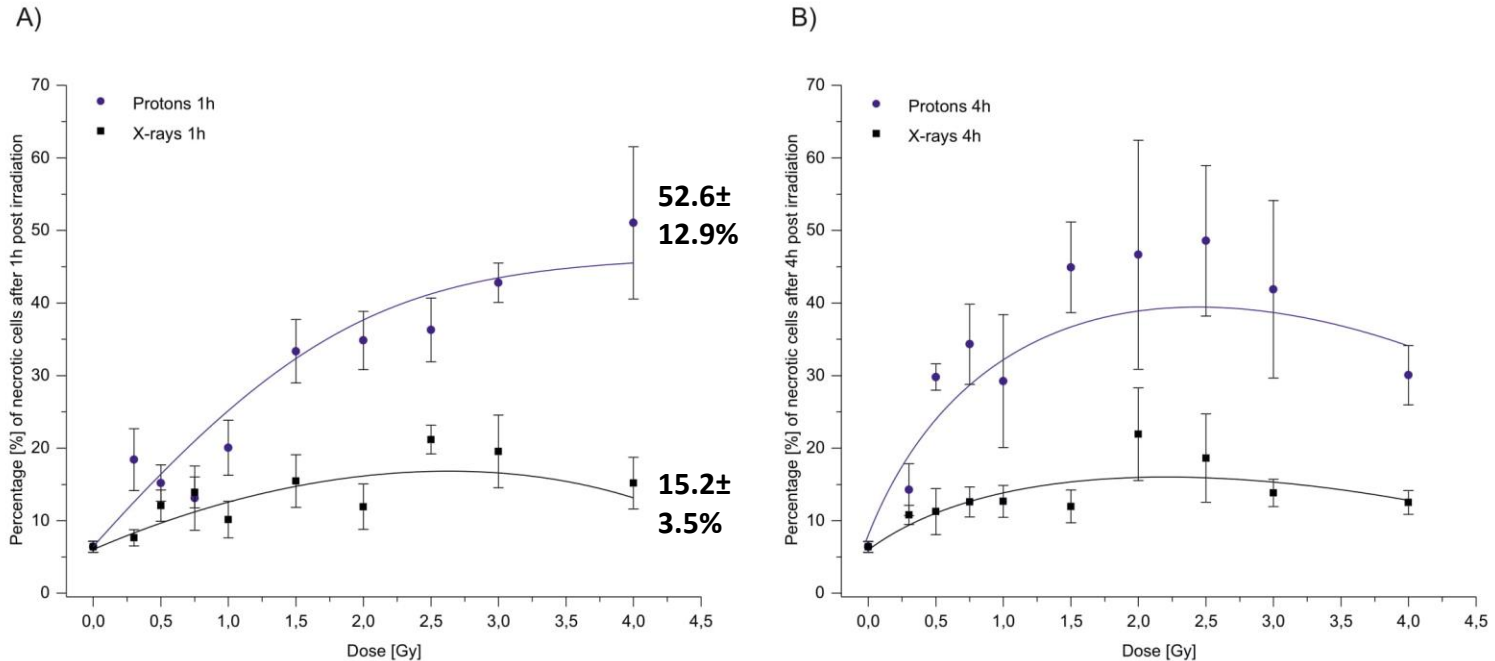


Fig. 23. Percentage of necrotic cells after irradiation with X-rays vs. protons (1 and 4 h after irradiation).



- ✓ **HPBLs, phantom, unique position.**
- ✓ **Easy, reliable biodosimetry protocols (PCC, CBMN) and different triage modes post proton and X-rays radiation for low and high dose exposure were proposed (possibility to discriminate whole and partial body exposures).**
- ✓ **Not only radiation (dose, type, dose-rate) but also the procedural steps determined the cell response to different degrees.**
- ✓ **Normal tissue response to protons vs. X-rays for low and high doses is different (influence cellular proliferation and DNA damage to different degrees).**

Effect depend on radiation type and dose.

Inter-individual differences determine response and effect.

Therefore, potentially influence the type, incidence and intensity of the RT AEs.



- ✓ Protons and X-rays induce cell-killing in normal tissue by different mechanisms. Protons are more efficient by necrosis. Inter-individual differences were also observed.
- ✓ Apoptosis is mediated by caspase-3, but necrosis?



Due to physical properties, protons and heavy ions deposit energy more selectively than

X-rays, allowing a higher local control of the tumor.

Thus, the damage induced in normal tissues surrounding the tumor is limited.

Protons are distinct from photons not only concerning their unique dosimetry but also with their ability to invoke unique biological responses that can be differentially exploitable.

Therefore, continued studies of these differences are necessary to benefit from a given type of radiation treatment.



Modeling Radioimmune Response – Current Status and Perspectives

Thomas Friedrich^{1*}, Nicholas Henthorn^{2,3} and Marco Durante^{1,4}

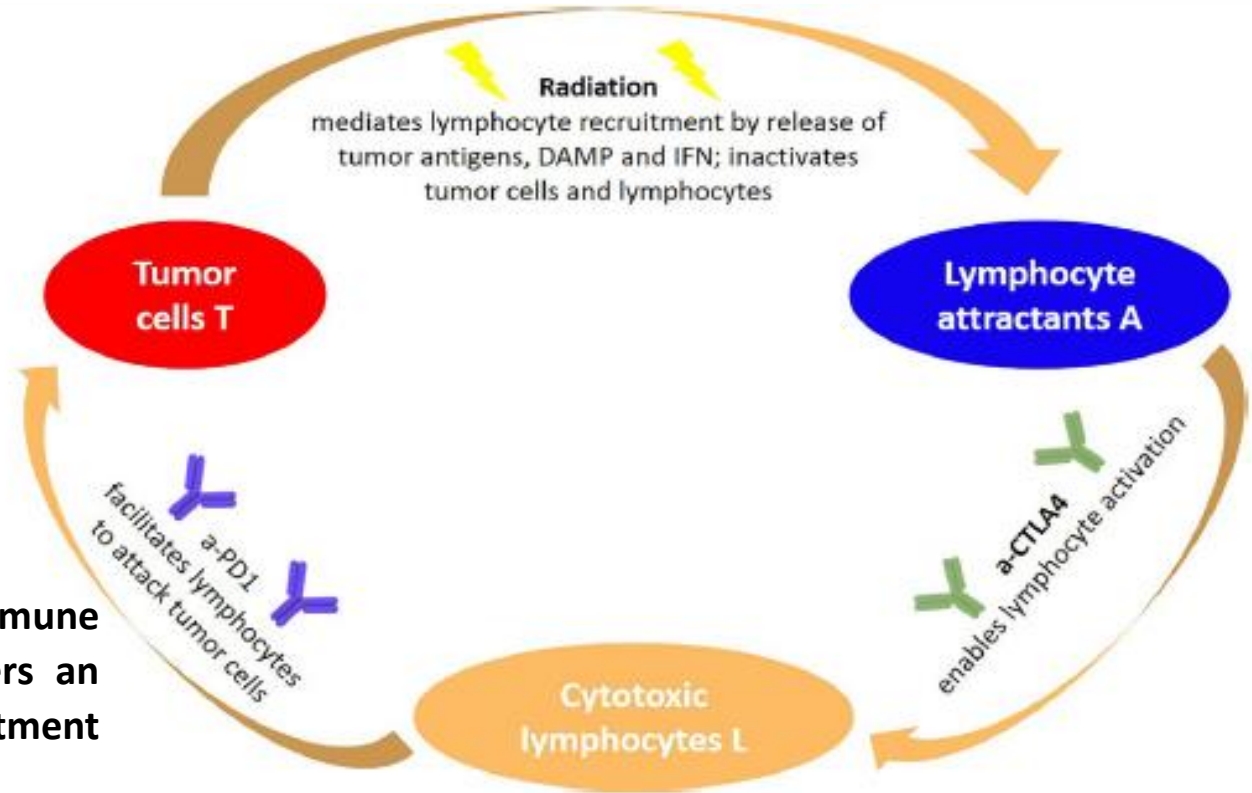


REVIEW

published: 16 March 2021

doi: 10.3389/fonc.2021.647272

Potential sensitizers
or radioprotectors can make tumor cells
more detectable for immune system.



General paradigm underlying RT using immune checkpoint blockers.

The combination of immune therapy with radiation offers an exciting and promising treatment modality in cancer therapy.



Cancer Treatment Reviews

Available online 16 April 2021, 102209

In Press, Journal Pre-proof



Anti-tumour Treatment

Proton beam therapy for children and adolescents and young adults (AYAs): JASTRO and JSPHO Guidelines

Masashi Mizumoto ^a, Hiroshi Fuji ^b, Mitsuru Miyachi ^c, Toshinori Soejima ^d, Tetsuya Yamamoto ^e, Norihiro Aibe ^f, Yusuke Demizu ^d, Hiromitsu Iwata ^g, Takayuki Hashimoto ^h,

The Japanese Society for Radiation Oncology (JASTRO)

The Japanese Society of Pediatric Hematology/Oncology (JSPHO).

- ✓ Careful use of PBT is recommended in adult patients, while sedation is required to maintain pediatric patients at rest.
- ✓ Further validation of irradiation techniques is needed for X-ray and PBT in both pediatric and adult patients.
- ✓ These guidelines show the superiority or equivalence of PBT in comparison with X-ray therapy for pediatric tumors.
- ✓ However, brainstem necrosis after PBT is still under discussion and requires further examination.

We need continue to evaluate the long-term efficacy of proton beam therapy for pediatric and adult patients.



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Agnieszka Burkat
Monika Gądek
Alina Kula
Katarzyna Magiera



Faculty of Physics
and Applied Computer Science

Irradiations

Cell's counted

~0.5 mln.

Image aquisition

Methods used

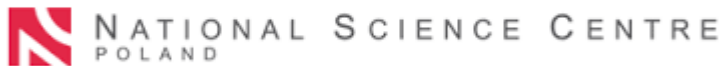
7; 12

Data were analyzed separately from
3 independent repetitions,
by min. 2 independent scorers.



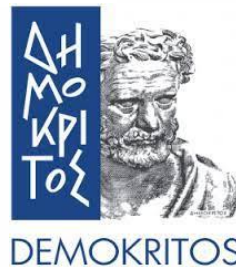
Laboratory of spectroscopic imaging for radiobiology, treatment and research of complex systems - equipment purchased as a part of the project co-funded by the Malopolska Regional Operational Program Measure 5.1 Krakow Metropolitan Area as an important hub of the European Research Area for 2007-2013.

Research supported by project DEC-2013/09/D/NZ7/00324 and POWR.03.05.00-00-Z309/17-00.



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Leiden University Medical Center,
Department of Toxicogenetics, Netherlands.



Federal Office for Radiation Protection (BfS), Germany

